

## Genetic diversity and conservation of Carex kobomugi in Taiwan

### Chi-Chun HUANG<sup>1,†</sup>, Chao-Yu HSU<sup>2,†</sup>, Kuo-Hsiang HUNG<sup>3,4</sup>, Tsai-Wen HSU<sup>5</sup>, Chau-Ching HUANG<sup>1</sup>, Munkhtsetseg TSEDNEE<sup>6</sup>, Chuan-Ming YEH<sup>7,8,\*</sup>, Wei-Kuang WANG<sup>9,\*</sup>

1. Wild Plants Division, Taiwan Biodiversity Research Institute, Nantou 552, Taiwan. 2. Division of Urology, Department of Surgery, Tungs' Taichung MetroHarbor Hospital, Taichung 435, Taiwan. 3. Graduate Institute of Bioresources, Pingtung University of Science and Technology, Pingtung 912, Taiwan. 4. Forestry and Biodiversity Research Center, National Pingtung University of Science and Technology, Pingtung 912, Taiwan. 5. Resource Management Division, Taiwan Biodiversity Research Institute, Nantou 552, Taiwan. 6. Research Center for Environmental Changes, Academia Sinica, Taipei 115, Taiwan. 7. Institute of Molecular Biology, National Chung Hsing University, Taichung 402, Taiwan. 8. Bioproduction Research Institute, National Institute of Advanced Industrial Science and Technology (AIST), Tsukuba, Ibaraki 305-8566, Japan. 9. Department of Environmental Engineering and Science, Feng Chia University, Taichung 407, Taiwan.<sup>†</sup> contributed equally to this work; \*Correspondence \*Corresponding authors' emails: CMY: chuanmingy@nchu.edu.tw; WKW: wkwang@fcu.edu.tw

(Manuscript received 21 November 2024; Accepted 6 June 2025; Online published 24 June 2025)

ABSTRACT: *Carex kobomugi* is native to the coastal sand dunes and sandy beaches of East Asia, including China, Japan, Korea, Taiwan, and the Russian Far East. In Taiwan, *C. kobomugi*, which is categorized as threatened species (category of critically endangered, CR), is narrowly distributed in northern Taiwan and offshore islands. First recorded in 1924, the species disappeared for a time but was rediscovered in 2020. However, it is currently threatened by increasing human activities and faces the risk of local extinction. In this study, we employed molecular markers (*rpL16* spacer of cpDNA, ITS region of nrDNA and eleven microsatellite loci) to evaluate the genetic diversity and population structure. The results showed that the low genetic diversity of *C. kobomugi* in Taiwan may be attributed to small population size, and geographic isolation based on cpDNA and nrDNA. In contrast to the identical nrDNA sequences across different regions, a unique cpDNA haplotype identified exclusively in Taiwan underscores the need for focused conservation efforts. A moderate level of genetic diversity was found for microsatellite loci, and STRUCTURE and principal coordinates analysis revealed that the two genetic groups corresponded to their geographical distributions. Based on these findings, two distinct management units are recommended for further conservation. Actions such as manually conducted beach cleanups, species inventories, and both *in-situ* and *ex-situ* conservation have been implemented to mitigate human disturbance.

KEY WORDS: Carex kobomugi, conservation, ITS, microsatellite, population genetics, rpL16.

## INTRODUCTION

Gene flow is an important force in maintaining species boundary by introducing genetic variation into populations and preventing population differentiation by local adaptation and genetic drift (García-Ramos and Kirkpatrick, 1997; Morjan and Rieseberg, 2004; Takayama et al., 2008; Sexton et al., 2011; M'Baya et al., 2013). Species with a high dispersal ability are expected to maintain species boundary through gene flow. In contrast, species with a low dispersal ability are more likely to diverge due to random mutations and genetic drift (Suárez et al., 2022). For coastal species, ocean currents play a crucial role in shaping their distribution (Schönswetter et al., 2008; Geng et al., 2021; Han et al., 2022). Specific morphological and ecophysiological adaptations can enable species to achieve long-distance dispersal via ocean currents (Takayama et al., 2006, 2008; Geng et al., 2021). In Taiwan, certain species, such as Kandelia candel (Chiang et al., 2001a) and Ipomoea pescaprae (Miryeganeh et al., 2014), have demonstrated that ocean currents play a crucial role in the long-distance dispersal of mangroves or seaside plants. Conversely, ocean currents can act as geographic barriers, hindering

gene flow between populations (Takayama *et al.*, 2006; Yamamoto *et al.*, 2019). Ochoa-Zavala *et al.* (2020) showed that the genetic discontinuities of *Avicennia germinans* along the coasts of Mexico were generated by ocean currents and a physical barrier. Ocean currents between the Andaman Sea and the Malacca Strait have been attributed to the distinct genetic discontinuity of *Rhizophora mucronata* (Wee *et al.*, 2014).

Taiwan, a continental island located approximately 150 km off the Southeast Asian coast, harbors significant species diversity (Chiang and Schaal, 2006). More than 4,000 taxa of vascular plants have been recorded in Taiwan (Hsieh et al., 1994). The island's coastal ecosystems are shaped by complex ocean currents, which play a crucial role in species distribution. The coastal community consists primarily of taxa with a Pacific Basin distribution, such as Kandelia obovata, I. pes-caprae, Artemisia capillaris, and Wollastonia biflora (Hsieh et al., 1994). However, long-term human activities have led to the fragmentation of coastal habitats, which has made it difficult to study the population genetics of seaside species. Chiang et al. (2001a) suggested that two differentiated lineages of K. candel corresponding to geographical regions. Low levels of genetic differentiation



between and within mainland and island populations indicated conspicuous long-distance seedling dispersal across oceans. Ruan *et al.* (2013) proposed that the population structure of *K. obovata* is affected by both different sources of colonizers and the geographic barrier. Long-term isolation by the Taiwan Strait differentiated the Taiwanese and Chinese populations. While mangroves have received more research attention (Chiang *et al.*, 2001a; Ruan *et al.*, 2013; Huang *et al.*, 2015), studies on other seaside plants remain limited (Huang and Hsu, 2023). Among these, *Carex kobomugi* Ohwi, a sand sedge found along Taiwan's coasts, was overlooked until 2020.

Carex L. (Cyperaceae), with about 2000 species, is one of the most species-rich genera among angiosperms (Ford et al., 2006). Carex kobomugi Ohwi is naturally distributed in the coastal sand dunes and sandy beaches of East Asia, including China, Japan, Korea, Taiwan, and Russian Far East (WFO, 2024). There are several studies on the phylogeny (Ford et al., 2006; King et al., 2009; King and Roalson, 2009), reproductive system (Ohsako, 2010), invasion (Charbonneau et al., 2020; Riffe and Zinnert, 2024). Carex kobomugi is closely related to C. macrocephala, which is naturally distributed in the coastal sand dunes and sandy beaches of North American, Russian and Japan (Ford et al., 2006; King and Roalson, 2008, 2009). King and Roalson (2009) adopted molecular data (chloroplast rpL16 and microsatellite DNA) to clarify the relationships of the C. macrocephala species complex, which includes the Asian C. macrocephala, North American C. macrocephala, and Asian C. kobomugi. The divergence among these lineages is estimated to have occurred during the Late Pleistocene epoch. Incomplete lineage sorting is considered the cause of the non-reciprocal monophyly observed among these lineages. Additionally, C. kobomugi was accidentally introduced to North America in 1929 and has become widespread along the Northeast coast of the USA. As an invasive species in the USA, efforts have been made to eradicate it (Wootton et al., 2005; Riffe and Zinnert, 2024).

In Taiwan, C. kobomugi, which is categorized as a threatened species (category CR: critically endangered) (Editorial Committee of the Red List of Taiwan Plants, 2017), is restricted to northern Taiwan and offshore islands. It was first recorded in 1924 and the specimen was deposited in NTU Herbarium (TAI no. 22663). However, few accurate records were reported until 2020. The known populations in Taiwan are only restricted to Fulon (FL, 福隆) and Mazu (MZ, 馬祖). The FL population, which has larger population size compared to MZ, was threatened by increasing human activities and was under the pressure of possible local extinction. A large amount of construction waste was dumped from 2011 to 2013 and was latter covered by sands, creating a habitat where most of C. kobomugi grows. In response to the impacts of habitat disturbance and human activities,

the conservation project of *C. kobomugi* has been ongoing since 2020. The construction waste is being manually removed to prevent machinery from affecting the population. However, limited knowledge about *C. kobomugi* in Taiwan complicates efforts to preserve the native populations. Specimen misidentification and incorrect information of sampling locations make it difficult to confirm the distributions of *C. kobomugi* in Taiwan. For known populations, urgent conservation measures, such as *in situ* preservation, germplasm collection and minimizing human disturbance, have been implemented to maintain the wild population size. The next critical question is whether these measures will be sufficient to preserve genetic diversity and structure.

Chloroplast DNA, internal transcribed spacer of nuclear ribosomal (nrDNA) and microsatellite DNA (simple sequence repeat, SSR) are widely used genetic markers for investigating phylogenetic relationships and population genetics (Ge et al., 2012; Feng et al., 2014; Liu et al., 2024; Wang et al., 2024). Different DNA markers have specific mutation rates, inheritance patterns, and dispersal mechanisms (Schlotterer, 2000; Kay et al., 2006; Huang et al., 2012). Chloroplast DNA evolves at a relatively slow rate and is maternally inherited, primarily reflecting the influence of seed dispersal on genetic diversity. In contrast, nrDNA, which evolves at a medium rate, and SSR, which evolves more rapidly, exhibit biparental inheritance, making them useful for assessing the impact of pollen flow on genetic diversity patterns (Feng et al., 2014; Liu et al., 2024). Furthermore, SSR serves as a valuable tool for analyzing the population structure of endangered species and providing essential information for conservation strategies (Wahlsteen, 2021; Hu et al., 2024; Ko et al., 2024). Species delimitation and identification of genetically distinct populations provide vital information for further effective conservation measures (Wang et al., 2024). In this study, we employed rpL16 (cpDNA), nrDNA and SSR markers to assess genetic diversity, population differentiation, and genetic structure, with the aim of informing conservation strategies.

## MATERIALS AND METHODS

#### **Plant materials**

Thirty-three individuals in Taiwan were sampled (Fig. 1, Table 1). Despite several field surveys conducted in the coastal region of northern Taiwan from 2020 to 2023, only two extent populations (FL and MZ) were confirmed. The habitat in FL has been partly covered by construction waste since 2011, leading to changes in the landscape. Although later buried by sand, the waste remains to this day. Based on personal observations, most *C. kobomugi* individuals in FL are found on the sand dunes above the waste, with an estimated population size of approximately 600 individuals. In contrast, the population size in MZ is estimated to be fewer than 100



**Table 1.** Sample locations, code, coordination, sample size, nucleotide diversity ( $\pi$ ), haplotype (h) and haplotype diversity (Hd) of **Carex kobomugi** in Taiwan.

Chaolea	Location	Codo	Longitudo	Latituda	Sample size	nrDNA			cpDNA		
Species	Location	ation Code Longitude Latitude Sample size		Sample size	π	h	Hd	π	h	Hd	
Carex koborr	Carex kobomugi				33	0	1	0	0	1	0
	Fulong	FL	121.937	25.029	18	0	1	0	0	1	0
	Mazu	MZ	119.981	25.966	15	0	1	0	0	1	0



Fig. 1. (A) The sample locations of *Carex kobomugi* in Taiwan, (B) graphical depiction of the relationship between K and  $\Delta K$ . (C) STRUCTURE histogram of genetic cluster membership generated from analysis of microsatellite loci for K =2–4. (D) Principal coordinates analysis (PCoA) of *C. kobomugi* based on microsatellite loci. FL: Fulon, MZ: Mazu.

individuals. Eighteen and fifteen individuals were collected form FL and MZ, respectively. Yong leaves were collected and dried using silica gels for genomic DNA extraction. The collected specimens were stored in TESRI herbarium.

#### DNA extraction, PCR amplification and sequencing

Genomic DNA was extracted from powdered tissues following the CTAB procedure (Murray andThompson, 1980). PCR amplification of the *rpL16* spacer (cpDNA; rpL16F71: GCTATGCTTAGTGTGTGACTCGTTG; rpL16R1516: CCCTTCATTCTTCCTCTATGTTG) (Shaw *et al.*, 2005) and ITS (nrDNA; ITS4: TCCTCCGCTTATTGATATGC; ITS5: GGAAGTAAAAGTCGTAACAAGG) (Chiang *et al.*, 2001b) was performed in a 50  $\mu$  L reaction using 10 ng template DNA, 25  $\mu$  L GoTaq ® Green Master Mix (Promega, Madison, WI, USA), and 5 pmol of each primer. PCRs was performed in a PCR cycler using the following profile: an initial 10 min denaturation at 94°C, 30 cycles of 45 s denaturation at 94 °C, 1 min 15 s annealing at 55 °C for two markers, and 1 min 30 s extension at 72 °C, followed by a 10 min final extension at 72 °C. All PCR products were purified and then sequenced directly in both directions on an ABI 3730XL automated sequencer (Applied Biosystems, Foster City, CA, USA). Direct sequencing of PCR products generates heterozygous base-calling fluorescence (double peak)

2025



chromatograms, which show paralogous genes within individuals (Chang *et al.*, 2012). In this study, the ITS sequences obtained from all individuals did not exhibit any double peaks for any of the sites in the chromatogram obtained by direct sequencing.

Eleven microsatellite loci (Ohsako and Yamane, 2007) were performed in a 25  $\mu$  L reaction using 5 ng template DNA, 12.5  $\mu$  L GoTaq ® Green Master Mix (Promega, Madison, WI, USA), and 2.5 pmol of each primer. PCRs was performed in a PCR cycler using the following profile: an initial 10 min denaturation at 94°C, 30 cycles of 45 s denaturation at 94 °C, 1 min of specific annealing temperature of each microsatellite locus, and 1 min extension at 72 °C. The PCR products were then analyzed using capillary electrophoresis on the 5200 Fragment Analyzer System (Agilent Technologies, USA), and allelic sizes were determined with the ProSize Data Analysis Software (Agilent Technologies, USA).

#### Data analysis

Sequences from National Center for Biotechnology Information (NCBI, https:// www. ncbi. nlm. nih. gov/; Supplementary table 1) were used as outgroups to infer the population genetics of Carex in Taiwan. For cpDNA and nrDNA, nucleotide sequences were aligned using MAFFT version 7 (Katoh et al., 2019) and later adjusted visually. The best-fit models of nucleotide substitution for the alignments were estimated with jModeltest 2.1.10 (Darriba et al., 2012). Maximum-likelihood (ML) analyses and Bayesian inference (BI) algorithms were performed to infer the relationships among the studied Carex haplotypes. ML analyses were performed using PhyML v. 3.67 (Guindon et al., 2010). The bootstrap consensus values were calculated using 1,000 replicates. BI tree was generated with the program MrBayes 3.2.7 (Huelsenbeck and Ronquist, 2001). Two independent Markov chain Monte Carlo (MCMC) runs with 5 X 10<sup>6</sup> generations were performed for the analysis. Trees were sampled every 1,000 generations. The first 25% of the sampled trees were discarded as burn-in, and the remaining trees were used to build a 50% majority-rule consensus tree. FigTree version 1.4.4 (Rambaut, 2018) and the "ggtree" package (Yu, 2020) were applied to depict the ML and BI trees. Levels of genetic diversity within species were quantified with measures of nucleotide diversity ( $\pi$ ) (Nei, 1987), haplotype number (h), haplotype diversity (Hd) using DnaSP 6 (Rozas et al., 2017). To depict relationship among Carex population belonging to different geographic regions, TCS network of cpDNA haplotype was conducted by PopART V.1.7 (Leigh and Bryant, 2015). CpDNA was used to estimate the divergence times. A mean mutation rate,  $\mu = 1 \times 10^{-1}$  $^{8}$  /site/year, with an interval set from 1  $\times$  10  $^{-7}$  /site/year to 1  $\times$  10  $^{-9}$  /site/ year, was used for the cpDNA data (Wolfe et al., 1987; Willyard et al., 2007). The

divergence time between Asian C. macrocephala and C. kobomugi (160,000 years with a 95% confidence interval of 75,000 to 296,000 years) was used as a calibration point (King and Roalson, 2009). We used BEAST version 2.6 to estimate the time of divergence with  $10^7$  iterations and a burn-in of 10<sup>6</sup> under the HKY model and a strict molecular clock (Bouckaert et al., 2019). TRACER v1.7.1 (Rambaut et al., 2018) was used to examine the convergence of chains to stationary distribution. TREEANNOTATOR v2.6.3 (Bouckaert et al., 2019) and FigTree version 1.4.4 (Rambaut, 2018) were used to summarize and display the sampled tree, respectively. For microsatellite DNA, to ensure the unique genotype, the multilocus genotypes (MLGs) were identified using R package "poppr" version 2.9.6 (Kamvar et al., 2014). The genetic diversity parameters, including the number of individuals (N), number of alleles (Na), the number of effective alleles (Ne), observed heterozygosity (Ho), expected heterozygosity (He), Shannon's information index (I), fixation index (F) were calculated using GenAlEx 6.51 (Peakall and Smouse, 2012). Deviations from Hardy-weinberg equilibrium in each population across all loci were tested using a chi-square test  $(X^2)$  in GenAlEx 6.51 (Peakall and Smouse, 2012). Polymorphic information content (PIC) was determined using Cervus 3.0.7 (Marshall et al., 1998). The results of analysis of molecular variance (AMOVA), and pairwise F<sub>ST</sub> analysis were performed to assess genetic differentiation within and among populations by ARLEQUIN 3.5.2.2 (Excoffier and Lischer, 2010). Significance tests were conducted using 10,000 permutations. Principal coordinates analysis (PCoA) was computed by GenAlEx 6.51 (Peakall and Smouse, 2012). Genetic structure was analyzed using a Bayesian clustering method implemented in STRUCTURE 2.3.4 (Pritchard et al., 2000). Independent simulations were run of each K (K =1-10) with 500,000 burn-in steps followed by 5,000,000 MCMC steps. Each analysis was repeat 5 times for each consecutive value of K. StructureSelector was utilized to visualized results from Structure (Li and Liu, 2018). BAYESASS software (Wilson and Rannala, 2003) that use a Bayesian approach and a Markov chain Monte Carlo (MCMC) algorithm was applied to estimate the recent migration rates between populations. We used 50,000,000 iterations, with a burn-in of 5,000,000 generations, and a sampling interval of 2000. The mixing parameters for migration rates (m), allele frequencies (a) and inbreeding coefficients (f) were optimized to achieve posterior acceptance rates of 20-60% for each parameter. Convergence of the MCMCs was checked by comparing the traces of each run using TRACER v. 1.7 (Rambaut et al., 2018). We conducted five independent runs with different random seed numbers to ensure consistent results.

Temporal changes in population size were estimated by the VarEff model, that use the coalescent method and



**Table 2.** Characteristics of the 11 polymorphic SSR markers usedin this study.

Locus	Ν	Na	Ne	1	Но	He	PIC	F
	32.909	6.545	3.890	1.414	0.586	0.669	0.633	0.122
Cko_1_9	33	11	7.358	2.118	0.545	0.864***	0.849	0.369
Cko_1_12	33	8	4.694	1.748	0.697	0.787***	0.759	0.114
Cko_1_47	33	12	7.003	2.168	0.545	0.857***	0.843	0.364
Cko_1_68	33	4	2.814	1.205	0.939	0.645***	0.599	-0.457
Cko_1_80	33	4	2.672	1.079	1.000	0.626***	0.554	-0.598
Cko_2_56	33	6	2.593	1.237	0.576	0.614***	0.577	0.063
Cko_2_112	33	6	4.605	1.636	0.939	0.783***	0.751	-0.200
Cko_2_113	33	3	2.562	1.005	0.000	0.610***	0.531	1.000
Cko_2_118	33	8	3.582	1.563	0.545	0.721***	0.688	0.243
Cko_2_135	33	3	1.130	0.263	0.091	0.115***	0.111	0.211
Cko_2_139	32	7	3.779	1.536	0.563	0.735***	0.697	0.235

N: number of individuals; *Na*: number of alleles; *Ne*: number of effective alleles; *I*: Shannon's information index; *Ho*: observed heterozygosity; *He*: expected heterozygosity; PIC: polymorphic information content; *F*: fixation index.

approximate likelihoods in a Monte Carlo Markov Chain approach (Nikolic and Chevalet, 2014). The VarEff method was implemented in the R package "VarEff"

Temporal changes in population size were estimated by the VarEff model, that use the coalescent method and approximate likelihoods in a Monte Carlo Markov Chain approach (Nikolic and Chevalet, 2014). The VarEff method was implemented in the R package "VarEff" (https://qgsp.jouy.inra.fr). This analysis was performed assuming a single step model, S, with a microsatellite mutation rate,  $\mu = 2 \times 10^{-4}$  (Escudero *et al.*, 2013), and burn-in of 10,000 over the past 10,000 generations. The parameter values included setting the assumed prior values for effective size to 10,000, number batch to 50,000, the length and space batch to 10, an acceptance rate of 0.25 and diagonale of 0.5, following recommendations from Nikolic and Chevalet (2014). The median of the posterior distribution was used to visualize demographic inference. Generation time was set to one, based on the fact that C. kobomugi is a perennial herb with annual reproduction (Ohsako, 2010).

To predict the suitable habitats for C. kobomugi, the maximum-entropy (MaxEnt) model was utilized (Phillips et al., 2006). The geographic distribution information of C. kobomugi was obtained from the data of Global Biodiversity Information Facility (GBIF, https://www.gbif.org/zh-tw/). 110 occurrence points from Japan, Korea, and Taiwan, recorded between 1970 and 2000, were used to compute species distribution models in MaxEnt. Nineteen bioclimatic factors and elevation data at 30 arc-second (~1km) resolution, for both current (1970-2000) and future (2081-2100, ACCESS-CM2, SSP 5-8.5) scenarios, were obtained from the WorldClim database v2.1 (https://www.worldclim.org/data/worldclim21.html). Bioclimatic factors corresponding to the species distribution were extracted and imported into "dismo" (Hijmans *et al.*, 2024) package in R. Strong correlations among environmental variables can reduce model accuracy due to multicollinearity. To address this, Pearson's correlation coefficients (r) were calculated for all pairs of variables and excluded one variable from each pair with  $|\mathbf{r}| > 0.8$  (Supplementary Fig. S1) (Rana *et al.*, 2017). Variables contributing more than 1% to model prediction results were initially retained (Dai *et al.*, 2022). Among these, the most influential environmental variables, based on contribution rates in the initial model were selected for further analysis.

Species distribution models were constructed using the MaxEnt function from the "dismo" package (Hijmans *et al.*, 2024). The model was run with 30 replicates under a cross-validation scheme, with 75% of the occurrence data randomly selected for training and the remaining 25% for testing in each replicate. The regularization multiplier was set to 1. The average value was used to calculate the potential geographic distribution. The accuracy of the operation results was evaluated using the ROC curve and the area under the curve (AUC) value. Values of AUC range from zero to one, with those closer to one indicating a more accurate model.

## RESULTS

# Genetic diversity of cpDNA, nrDNA and microsatellite loci

The levels of cpDNA (*rpl16*) and nrDNA genetic diversity of *Carex kobomugi* in Taiwan were examined, as showed in Table 1. The consensus sequences for cpDNA and nrDNA were 756 and 646 bp in length, respectively. No polymorphic sites were detected in either cpDNA (GenBank accession numbers: PQ351396) or nrDNA (GenBank accession numbers: PQ345421), indicating that individuals in both regions were fixed in the single haplotype. Nucleotide ( $\pi$ ) and haplotype (*Hd*) diversity were both found to be zero in cpDNA and nrDNA.

Genetic diversity was assessed in 33 individuals from two C. kobomugi populations using 11 microsatellite markers (Table 2). Allele size ranged from 157 bp (Cko 1 47) to 293 bp (Cko 2 118), with a total of 72 alleles across all loci. The number of alleles per locus (Na) ranged from 3.000 (Cko\_2\_135) to 12.000 (Cko\_1\_47), with a mean value of 6.545. The number of effective alleles (Ne) varied from 1.130 (Cko 2 135) to 7.358 (Cko 1 9), with an average of 3.890. Observed (Ho) and expected (He) heterozygosity ranged from 0 to 1.000 and 0.115 to 0.864, with a mean values of 0.586 and 0.669, respectively. The polymorphic information content (PIC) ranged from 0.111 to 0.849, with a mean value of 0.633. A total of 33 multilocus genotypes (MLGs) were identified from 33 samples using the "poppr" package in Significantly deviation from Hardy-Weinberg R. equilibrium (HWE) were detected in all of the loci.

The levels of genetic diversity in the two populations



Fig. 2. Phylogenetic tree of nrDNA (A) and cpDNA (B) based on a maximum-likelihood analysis. The numbers on the branches are bootstrap values (>60)/posterior probabilities (>0.6). Sequence obtained in this study is shown in red, and those retrieved from NCBI are presented in regular black text. ACM: Asian *Carex macrocephala*, CK: *Carex kobomugi*. Sequence obtained in this study is shown in red, and those retrieved from NCBI are presented in regular black text.

Table 3. Genetic diversity of Carex kobomugi in Taiwan.

Species Location	Ν	Na	Ne	1	Но	Не	F
Carex kobomugi	32.909	6.545	3.890	1.414	0.586	0.669	0.122
FL	18	5.273	3.713	1.327	0.833	0.668	-0.220
MZ	15	3.273	2.316	0.831	0.285	0.461	0.457
N: number of indi	viduals:	Na: n	umber	of all	eles: A	<i>le</i> : nur	nber of

effective alleles; *I*: Shannon's information index; *Ho*: observed heterozygosity; *He*: expected heterozygosity; *F*: fixation index. FL: Fulon; MZ: Mazu.

**Table 4.** Analysis of Molecular Variance (AMOVA) based on SSR loci for Carex kobomugi.

Source of variation	df	Sum of squares	Variance components	Percentage variation
Between populations	1	33.3417	0.92089	21.93
Within populations	64	209.856	3.27899	78.07
Total	65	234.273	4.19988	100

are shown in Table 3. *Na* ranged from 3.273 to 5.273, while *Ne* varied from 2.316 to 3.713. *Ho* and *He* ranged from 0.285 to 0.833 and 0.461 to 0.668, respectively. *F* varied from -0.220 to 0.457, and number of private alleles ranged from 1.273 to 3.273, respectively. Higher genetic diversity was detected in FL population compared to MZ.

#### **Genetic differentiation**

A single nrDNA haplotype (PQ345421) was found in Taiwan, identical to NCBI samples from Japan (EU001022), South Korea (JX644826), and the USA (MN761780) (Fig. 2A). In contrast, a higher level of genetic differentiation in cpDNA was detected between Taiwan (PQ351396) and others countries (Taiwan vs. Japan (FJ424683–FJ424685, FJ424688–FJ424690),  $F_{ST}$ = 0.746; Taiwan vs. South Korea (FJ424686–FJ424687) 422

#### $F_{ST} = 0.889$ ) (Fig. 2B).

For microsatellite loci, pairwise  $F_{ST}$  values and AMOVA were conducted to evaluate genetic variation between two *C. kobomugi* populations in Taiwan (Table 4). The genetic differentiation between FL and MZ was moderate ( $F_{ST} = 0.200$ , p < 0.05), indicating a moderate genetic differentiation between two populations. The AMOVA results indicated that 79.97 % of the total genetic variation occurred within populations and 21.93% was attributed to differences between populations.

The BAYESASS was applied to estimate the recent migration rates between populations based on microsatellite DNA. The rate of gene flow from FL population to MZ population was 0.017 (95% CI = 0.0004-0.0586), while the rate in the opposite direction was 0.019 (95% CI = 0.0005-0.0683). In contrast, the rates of gene flow within FL and MZ populations were 0.983 95% (CI = 0.9414-0.9996) and 0.981 (95% CI = 0.9317-0.9995), respectively.

### Phylogenetic reconstruction and Genetic structure

The nrDNA and cpDNA phylogenetic trees, reconstructed using both ML and BI methods, revealed consistent topologies. Therefore, only the ML trees are presented, with Bayesian posterior probabilities indicated at the corresponding nodes. (Fig. 2). The HKY+G model, identified as the most suitable model by jModeltest, was used to construct the phylogenetic trees for nrDNA and cpDNA. *Carex preslii* was set as outgroup for nrDNA, while *C. olbiensis* was used as outgroup for cpDNA.

For nrDNA (Fig. 2A), Clade A contains haplotypes of *C. kobomugi* (PQ345421 from Taiwan is identical to NCBI sequences from Japan (EU001022), the USA (DQ115194, MN761780), and South Korea (JX644826)).





Fig. 3. Results of coalescence time estimations performed using BEAST, v 2.6. The numbers at each nodes and in parentheses represent the estimated divergence times (in Mya) and their corresponding 95% highest posterior density (HPD) intervals. Sequence obtained in this study is shown in red, and those retrieved from NCBI are presented in regular black text.

Clade A is closely related to clade B, which consisted of all *C. macrocephala* (AF285017, EU001029, EU001030, and MN761779).

For cpDNA (Fig. 2B), *C. kobomugi* formed a monophyletic clade (Clade CK), which comprised subclade I (Japan: FJ424683–FJ424684, FJ424688–FJ424690), II (South Korea: FJ424686–FJ424687), III (Japan: FJ424685) and IV (Taiwan: PQ351396). Subclade IV was distantly related to others. Clade CK was closely related to clade ACM (Asian *C. macrocephala*: FJ424691–FJ424699). *Carex divulsa* (OZ060211), *C. arenaria* (OZ060148) and *C. obliensis* (MK970806) were used as outgroups.

Bayesian estimates of the divergence time of *C. kobomugi* based on cpDNA were obtained using the BEAST program (Fig. 3). The Taiwan lineage (PQ351396) was the first to diverge from the other lineages, with an estimated divergence time of approximately 0.11 may (95% HPD: 0.06–0.16).

The TCS network of cpDNA haplotypes revealed thatthe clustering corresponded with geographical distribution, with samples from different regions forming distinct clades. The haplotype from Taiwan was unique and distinct from those in Japan and South Korea (Fig. 4).

The STRUCUTRE results revealed that  $\Delta K$  reached a maximum value when K = 2 (Fig. 1), indicating that the

33 samples can be divided into two subgroups. The subgroup membership probabilities of 33 individuals from the two populations are shown in Fig. 1. These two subgroups corresponded to their geographical distribution. Also, the PCoA results (Fig. 1) were consistent with the STRUCTURE results.

VarEff was used to estimate the current and past effective population sizes and the changing times in *C. kobomugi* (Figs. 5 & S2). The results showed that the effective population size remained stable until approximately10,000 years ago, after which it began to decline, assuming a generation time of one year and a fixed mutation rate of  $2 \times 10^{-4}$  (Ohsako, 2010; Escudero *et al.*, 2013). Different populations (FL and MZ) experienced different demographic histories. FL underwent recent population expansion following a dramatic decline, maintaining a larger population size, while MZ experienced population decline and remained smaller.

# Distribution of suitable habitats for C. kobomugi in Taiwan

The MaxEnt model for *C. kobomugi* had high predictive power (mean AUC value = 0.994) for inferring the suitable habitats in Taiwan (Fig. 6). Based on MaxEnt modeling results, Precipitation of Warmest Quarter (bio18; 27.6%), Elevation (26.1%) Precipitation of Driest



Fig. 4. The TCS network of *Carex macrocephala* and *C. kobomugi* based on cpDNA haplotype. Each circle represents a haplotype, and the circle size is proportional to the haplotype frequency. ACM: Asian *C. macrocephala*, CK: *C. kobomugi* (KR: South Korea, JP: Japan, TW: Taiwan).



Fig. 5. Population size fluctuation of *Carex kobomugi.*  $\mu$ , mutation rate per locus per generation. FL: Fulon, MZ: Mazu.



Fig. 6. ROC curves in the developing *Carex kobomugi* distribution model. The red (training) shows the fit of the model to the training data. The blue (testing) line indicates the fit of the model to the testing data.

Month (bio14; 21.0%), and Temperature Seasonality (bio4; 18.9%) accounted for over 93.6% of the contribution (Supplementary table S2). These results indicates that precipitation, temperature variation and elevation are critical factors influencing the suitability of C. kobomugi (Fig. 7). We selected four variables to predict the current and future distributions for C. kobomugi (Figs. 8 & S3). The current potential distribution areas of C. kobomugi are located in the coastal regions of northern Taiwan and match the observed distributions. The future potential areas of C. kobomugi in Taiwan are contracted to several suitable habits scattered in the current distribution range. Environmental variables differed between FL and MZ, including Precipitation of Warmest Quarter (708 mm vs. 372 mm), Precipitation of Driest Month (179 mm vs. 41 mm), Temperature Seasonality (466.439 vs. 648.710) and Elevation (18 m vs. 30 m).

### DISCUSSION

#### Genetic diversity

CpDNA and nrDNA, which have been widely used for phylogenetic analyses of *Carex* (Roalson and Friar, 2004; King and Roalson, 2008, 2009; Yano *et al.*, 2010), were collaboratively applied to study the genetic diversity of *Carex* in Taiwan. For *C. hakkodensis* Franch, *C. scita* Maxim and *C. stenatha* Franch. Et Sav., only one to four cpDNA haplotypes were identified in Japan (Senni *et al.*, 2005), while a higher number of cpDNA haplotypes were detected in *C. conica* Boott (Yano *et al.*, 2010), *C. macrocephala* and *C. kobomugi* (Japan and South Korea) (King *et al.*, 2009). In this study, a single haplotype was found for both cpDNA and nrDNA, revealing that *C. kobomugi* in Taiwan possessed low genetic diversity (Table 1). *Carex kobomugi*, which is primarily distributed





Fig. 7. Response curves (blue indicates standard deviation) for important environmental predictors in MaxEnt model for Carex kobomugi:



Fig. 8. Potential distributions of *Carex kobomugi*. (A) Present (1970–2000) and (B) future potential distribution (2081–2100) were predicted using MaxEnt based on four bioclimatic variables.

in temperate Asia, has a peripheral distribution in Taiwan. Peripheral populations are often located in less optimal environments are isolated from and central and continuous populations. The smaller population size of peripheral populations make them more vulnerable to stochastic or catastrophic demographic events (Lammi et al., 1999; Lesica and Allendorf, 1995). Peripheral populations are expected to exhibit low genetic diversity (Channell, 2004). The lack of genetic diversity in C. kobomugi in Taiwan may be attributed to suboptimal habitats, small population sizes and prolonged isolation from other populations (Madsen et al., 1999; Furlan et al., 2012).

Different molecular markers are used to assess genetic diversity, population structure, and phylogeographical patterns. Microsatellite loci provide more detailed information on genetic structure, while cpDNA and nrDNA sequences are more effective for detecting phylogeographical patterns (Zhang et al., 2020, 2021; Liu et al., 2024). Microsatellite loci have been widely applied to evaluate the genetic diversity of Carex species, such as C. breviculmsi (Ho = 0.057), C. hebes (Ho = 0.105) (M'Baya et al., 2013), C. macrocephala (Ho = 0.041) (King et al., 2009) and C. kobomugi (Ho = 0.575–0.658) (Ohsako, 2010).

A total of 33 multilocus genotypes (MLGs) were identified among 33 samples in Taiwan. Our analysis of *C. kobomugi* from two populations revealed significant deviations from HWE due to heterozygote deficiency (table 2). King *et al.* (2009) proposed that high levels of selfing (F = 0.979) contributed to departures from HWE in *C. macrocephala*. Similarly, the positive *F* value (F =0.122) observed in *C. kobomugi* suggests nonrandom mating led to deviations from HWE (table 3).

A lower level of heterozygosity (Ho = 0.586) was observed compared to that in Japan (Ho = 0.575-0.658) at the same loci (Ohsako, 2010). The low levels of genetic diversity in cpDNA, nrDNA and SSR suggested that *C*. *kobomugi* in Taiwan corresponds to peripheral populations, which tend to occur in less suitable habitats and are often smaller, making them more prone to extinction due to stochastic or catastrophic demographic events.

However, different populations have experienced different population dynamics. Peripheral populations often exhibit genetic and morphological divergence from central populations (Lesica and Allendorf, 1995). In the FL population, higher observed heterozygosity (Ho = 0.833) and a negative inbreeding coefficient (F = -0.220) were detected, whereas the MZ population showed a lower observed heterozygosity (Ho = 0.285) and a positive inbreeding coefficient (F = 0.457). Geographic isolation, decreased gene exchange, and small population sizes increase the risk of genetic drift and inbreeding (Ellstrand, 1993; Madsen *et al.*, 1999; Ko *et al.*, 2024). The VarEff results (Fig. 5) showed that the FL population

underwent recent population expansion following a dramatic decline, maintaining a larger population size, while the MZ population experienced population decline and remained smaller. The FL population, characterized by a larger population size and a tendency for outcrossing, displayed greater genetic diversity. Ohsako (2010) proposed that the prolonged lifespan of individuals and mating system of dioecy could contribute to the maintenance of genetic diversity. In contrast, the MZ population, with its smaller size and inclination toward inbreeding, exhibited lower genetic diversity.

# Phylogeographic pattern, genetic differentiation and genetic structure

King and Roalson (2009) proposed that the divergence among Asian C. macrocephala, North American C. macrocephala, and C. kobomugi occurred during the Late Pleistocene epoch based on *rpL16* spacer of cpDNA and microsatellite loci. The estimated divergence time between Asian C. macrocephala and C. kobomugi is approximately 0.16 mya. Most of Taiwan' flora is thought to have originated during the Pleistocene glacial cycles (Chen et al., 2023). The single nrDNA haplotype from Taiwan was identical to NCBI sequences from Japan, the USA, and South Korea, which is consistent with King and Roalson's finding that nuclear markers show little to no variation within Carex (King and Roalson, 2008). In contrast, the single cpDNA haplotype from Taiwan was distantly related to those from other regions. In this study, the phylogeographic pattern of C. kobomugi was assessed using the cpDNA. Based on the BEAST results, all C. kobomugi haplotypes coalesced approximately 0.11 mya, and PQ351396 (the Taiwan-inclusive haplotype) diverged from the rest, suggesting that C. kobomugi migrated into Taiwan during the Late Pleistocene. Furthermore, based on microsatellite loci, the VarEff results showed that C. kobomugi maintained a stable population size but experienced a recent population decline after the Last Glacial Maximum. During the Pleistocene glacial period, the retreat of coastlines on both sides of the Taiwan Strait led to the disappearance of mangrove habitats in the region (Ruan et al., 2013). Consequently, habitat degradation following the Last Glacial Maximum may have contributed to the population decline of C. kobomugi. The persistently small effective population size observed in MZ suggests that MZ has long experienced suboptimal environmental conditions, possibly due to the loss of suitable coastal habitats during the Last Glacial Maximum. In contrast, the population decline in FL appears to have occurred more recently, likely as a result of habitat loss caused by post-glacial sea-level rise.

For both nrDNA and cpDNA, a single haplotype was detected in Taiwan populations, indicating the lack of genetic differentiation between populations. However, the genetic structure of *C. kobomugi* was found to consist



of two distinct groups: FL and MZ, as revealed by the STRUCTURE and PCoA of microsatellite loci. The  $F_{ST}$  (0.200) also suggested moderate genetic differentiation between the FL and MZ populations. The study revealed genetic difference in population structures between FL and MZ populations. Two possible explanations could account for this pattern.

First, the higher contemporary gene flow within populations (m = 0.980 - 0.983) were detected than that between populations (m = 0.017 - 0.020). These results correspond with the central-marginal hypothesis that peripheral populations have lower genetic diversity and higher genetic similarity because of genetic drift and higher local gene flow (Eckstein et al., 2006). Takayama et al. (2008) proposed that the high differentiation observed in African populations of Hibiscus tiliaceus was due to the geographical barrier of the African continent, whereas the differentiation in *H. pernambucensis* was attributed to the bifurcating South Equatorial Current. Yamamoto et al. (2019) suggested that ocean currents could act as cryptic barriers, corresponding to the population differentiation observed in Vigna marina. The buoyant seeds of C. kobomugi are capable of sea-water dispersal (Ishikawa et al., 1993; Yang et al., 2012). Ishikawa et al. (Ishikawa et al., 1993) proposed that the moist-chilling condition in winter help C. kobomugi seeds germinate in the following spring. The southward current, the Zhejiang-Fujian Coastal Current, the northward Taiwan Warm Currents, and the incursion of the Kuroshio Current branch, control the hydrological conditions in the Taiwan Strait (Liao et al., 2018). The combined effects of these currents may act as a geographic barrier, hindering gene flow between populations. Therefore, differentiation between the Carex populations might have been caused by successful colonization by sea-drifted seeds followed by an interruption of gene flow.

Second, local adaptation may shape the population structure (Savolainen et al., 2013). Selective factors, such as climate, edaphic factors and parasites can contribute to local adaption, and evolved traits can respond to selection pressures (Savolainen et al., 2007). Compared to genetic drift, which leads to random population structure changes (Yan et al., 2022), local adaption may drive the structure towards a specific composition based on selective pressures. Fournier-Level et al. (2011) proposed that the fitness-associated loci of Arabidopsis thaliana exhibited local adaptation with geographic and climatic signatures. Fan et al. (Fan et al., 2023) proposed that the correlation between genetic variation in microsatellite markers and climatic variables is stronger than that with geographic variables, suggesting the climatic factors are the primary drivers of local adaptation of Toona ciliata var. pubescens. Based on the worldclim2 data, environmental variables (Precipitation of Warmest Quarter and Driest Month and Temperature Seasonality) differed between FL and MZ.

These differences in climate and environmental factors could drive the FL and MZ populations into distinct genetic structure, leading to local adaption. Further studies using correlation analysis between morphological traits and genomic data are recommended for future research (Zimmerman *et al.*, 2020).

SDM analysis indicated that Precipitation of Warmest Quarter, Precipitation of Driest Month, Temperature Seasonality and Elevation contributed the most to model development. Based on the current potential distribution areas (Fig. 8), Taiwan represents a less suitable habitat, which is consistent with hypothesis of peripheral populations. The contracted habitats in the future potential areas implies that *C. kobomugi* in Taiwan is in critical situation and requires conservation efforts.

## Conservation strategies of Carex kobomugi in Taiwan

Maintaining higher genetic diversity, adaptive potential and avoiding inbreeding depression are crucial for conservation (Kardos et al., 2021). The cpDNA results suggest that the haplotype in Taiwan is distinct from those in Japan and South Korea. Two possible explanations could account for this: First, populations in Taiwan, located at the southern boundary of the species' range, may have a different evolutionary history compared to northern populations. One lineage may have split from the northern populations, migrated to Taiwan, and gradually undergone lineage sorting. Second, only limited genetic studies have focused on C. kobomugi (Ford et al., 2006; King et al., 2009; King and Roalson, 2009; Ohsako, 2010). The haplotype found exclusively in Taiwan could potentially be present elsewhere but has yet to be discovered due to insufficient sampling. Currently, the eight cpDNA haplotypes available from the NCBI database constitute the only genetic evidence used for this study.

The identification of Management Units (MUs) provide essential information for conservation. Preserving MUs, which are populations with significant genetic divergence, helps maintain the species diversity (Palsbøll et al., 2007; Coates et al., 2018). Carex kobomugi, with only two populations and small population size in Taiwan, has been categorized as Critically Endangered (CR) (Editorial Committee of the Red List of Taiwan Plants, 2017). Recent increases in human activities may lead to habitat loss. Microsatellite loci results suggest that FL and MZ exhibit different genetic structures and high percentage of variation (78.07 %) within populations, indicating that the two populations should be considered independent units. Furthermore, FL shows higher genetic diversity and a larger population size, making it a priority for conservation efforts.

Conservation efforts for *C. kobomugi* in Taiwan were initiated after its rediscovery in 2020. Actions such as manually conducted beach cleanups (replacing machinery), species inventories, and both *in-situ* and *ex*-



situ conservation have been implemented to mitigate human disturbance. Clugston *et al.* (2024) employed RADSeq data to confirm the uniqueness of *Sabal antillensis* and *S. lougheediana*, proposing further conservation implications and recommendations to protect these species. This study confirmed the presence of a unique cpDNA haplotype in Taiwan, and both populations exhibited distinct genetic components, suggesting that each should be considered in further conservation strategies. The genetic variation of *C. kobomugi* exists mainly within populations, and the conservation efforts to maintain the genetic diversity within populations are suggested.

First, to protect the existing populations, meetings with authority in charge were held and management strategies were modified to reduce human disturbances. For example, construction waste is being manually removed to prevent machinery from impacting the population. Second, germplasm collections from two populations are essential for preserving genetic diversity. During our field observations of the FL population in 2023-2024, approximately 600 individuals were recorded and seed production was found to be low, restricting the potential for seed-based reproduction in exsitu conservation. Therefore, asexual reproduction via cutting propagation is required to maintain the population size. Identifying the MLGs of C. kobomugi for asexual reproduction helps sustain higher genetic diversity. Currently, genetic materials from both populations have been collected and ex-situ preserved at TBRI (Taiwan Biodiversity Research Institute). Carex kobomugi is a perennial herb that can propagate sexually via wind pollination and asexually via elongating rhizomes (Ohsako, 2010). The identification of different MLGs using SSR markers, along with continued germplasm collection, is recommended to enhance ex-situ conservation efforts.

Global climate change, which causes the rise in sea level, poses a significant threat to coastal plants (Garner *et al.*, 2015). Inundation, flooding, and erosion, which are caused by sea level rise, could lead to habitat loss. This study provides insights into the genetic conservation and population dynamics of *C. kobomugi*. We identified distinct management units and collected genetic materials for *ex-situ* preservation. These actions not only help preserve genetic diversity but also contribute to the conservation of *C. kobomugi* in Taiwan. Our comprehensive approach aims to ensure the survival of *C. kobomugi* through appropriate management strategies.

## CONCLUSION

In summary, this study used molecular markers to assess the genetic diversity, population differentiation and genetic structure of *C. kobomugi*. As peripheral populations in Taiwan, *C. kobomugi* exhibits a unique haplotype compared to those from Japan and Korea. A distinct evolutionary history and geographic isolation have led to the genetic differentiation between the FL and MZ populations. Based on these findings, two distinct management units are recommended for further conservation efforts. Actions such as manually conducted beach cleanups, species inventories, and both *in-situ* and *ex-situ* conservation have been implemented to mitigate human disturbance.

## ACKNOWLEDGMENTS

This research was supported by the National Science and Technology Council, Taiwan, grant number NSTC 112-2221-E-035-007-MY3. We thank the reviewers for providing comments that improved the manuscript.

## LITERATURE CITED

- Bouckaert, R., Vaughan, T.G., Barido-Sottani, J., Duchêne, S., Fourment, M., Gavryushkina, A., Heled, J., Jones, G., Kühnert, D., DeMaio, N., Matschiner, M., Mendes, F.K., Müller, N.F., Ogilvie, H.A., DuPlessis, L., Popinga, A., Rambaut, A., Rasmussen, D., Siveroni, I., Suchard, M., Wu, C.H., Xie, D., Zhang, C., Stadler, T., Drummond, A.J. 2019 BEAST 2.5: An advanced software platform for Bayesian evolutionary analysis. PLoS Comput. Biol. 15(4): e1006650.
- Chang, C.T., Tsai, C.N., Tang, C.Y., Chen, C.H., Lian, J.H., Hu, C.Y., Tsai, C.L., Chao, A., Lai, C.H., Wang, T.H., Lee, Y.S. 2012 Mixed sequence reader: a program for analyzing DNA sequences with heterozygous base calling. Sci. World J. 2012(1): 365104.
- Channell, R. 2004 The conservation value of peripheral populations : the supporting science. In: T. D.Hooper (eds.), Proceedings of the species at risk 2004 pathways to recovery conference. 1–17. Species at Risk 2004 Pathways to Recovery Conference Organizing Committee, Victoria, BC.
- Charbonneau, B.R., Nicoletta, R., Wootton, L.S. 2020 A decade of expansion of the invasive plant *Carex kobomugi* in a coastal foredune system. Biol. Invasions **22(6)**: 2099–2112.
- Chen, S.Y., Huang, C.C., Cheng, Y.T., Wang, C.C., Li, C.Y., Lai, I.L., Hung, K.H. 2023 Effect of geographic isolation on genetic variation and population structure of *Euphrasia nankotaizanensis*, a threatened endemic alpine herb in Taiwan. Heliyon 9(3): e14228.
- Chiang, T.Y., Chiang, Y.C., Chen, Y.J., Chou, C.H., Havanond, S., Hong, T.N., Huang, S. 2001a Phylogeography of *Kandelia candel* in East Asiatic mangroves based on nucleotide variation of chloroplast and mitochondrial DNAs. Mol. Ecol. 10(11): 2697–2710.
- Chiang, T.Y., Hong, K.H., Peng, C.I. 2001b Experimental hybridization reveals biased inheritance of the internal transcribed spacer of the nuclear ribosomal DNA in *Begonia* ×*taipeiensis*. J. Plant Res., **114(3)**, 343–351.
- Chiang, T.Y., Schaal, B.A. 2006 Phylogeography of plants in Taiwan and the Ryukyu Archipelago. Taxon 55(1): 31–41.
- Clugston, J.A.R., Coolen, Q., Houtepen, E., Van Proosdij, A. S.J., Grinage, A.D., Griffith, M.P. 2024 Genomic patterns of native palms from the Leeward Antilles confirm single-



island endemism and guide conservation priorities. Conser. Genet. **25(4)**: 985–997.

- Coates, D.J., Byrne, M., Moritz, C. 2018 Genetic diversity and conservation units: Dealing with the species-population continuum in the age of genomics. Front. Ecol. Evol. 6: 165.
- Dai, X., Wu, W., Ji, L., Tian, S., Yang, B., Guan, B., Wu, D. 2022 MaxEnt model-based prediction of potential distributions of *Parnassia wightiana* (Celastraceae) in China. Biodivers. Data J. 10: e81073.
- Darriba, D., Taboada, G.L., Doallo, R., Posada, D. 2012 JModelTest 2: More models, new heuristics and parallel computing. Nat. Methods 9(8): 772.
- Eckstein, R.L., O'Neill, R.A., Danihelka, J., Otte, A., Köhler, W. 2006 Genetic structure among and within peripheral and central populations of three endangered floodplain violets. Mol. Ecol. 15(9): 2367–2379.
- Editorial Committee of the Red List of Taiwan Plants 2017 The Red List of Vascular Plants of Taiwan, Endemic Species Research Institute, Forestry Burea. 187 pp.
- **Ellstrand**, N.C. 1993 Population genetic consequences of small population size: implications for plant conservation. Annu. Rev. Ecol. Syst. **24(1)**: 217–242.
- Escudero, M., Weber, J.A., Hipp, A.L. 2013 Species coherence in the face of karyotype diversification in holocentric organisms: the case of a cytogenetically variable sedge (*Carex scoparia*, Cyperaceae). Ann. Bot. 112(3): 515–526.
- Excoffier, L., Lischer, H.E.L. 2010 Arlequin suite ver 3.5: A new series of programs to perform population genetics analyses under Linux and Windows. Mol. Ecol. Resour., 10(3): 564–567.
- Fan, Y., Dai, J., Wei, Y., Liu, J. 2023.Local adaptation in natural populations of *Toona ciliata* var. *pubescens* is driven by precipitation and temperature: evidence from microsatellite markers. Forests 14(10): 1998.
- Feng, X., Wang, Y., Gong, X. 2014 Genetic diversity, genetic structure and demographic history of *Cycas simplicipinna* (Cycadaceae) assessed by DNA sequences and SSR markers. BMC Plant Biol. 14(1): 187.
- Ford, B.A., Iranpour, M., Naczi, R.F.C., Starr, J.R., Jerome, C.A. 2006 Phylogeny of *Carex* subg. *Vignea* (Cyperaceae) based on non-coding nrDNA sequence data. Syst. Bot. 31(1): 70–82.
- Fournier-Level, A., Korte, A., Cooper, M.D., Nordborg, M., Schmitt, J., Wilczek, A.M. 2011 A map of local adaptation in Arabidopsis thaliana. Science, 334(6052): 86–89.
- Furlan, E., Stoklosa, J., Griffiths, J., Gust, N., Ellis, R., Huggins, R.M., Weeks, A.R. 2012 Small population size and extremely low levels of genetic diversity in island populations of the platypus, *Ornithorhynchus anatinus*. Ecol. and Evol. 2(4): 844–857.
- García-Ramos, G., Kirkpatrick, M. 1997 Genetic models of adaptation and gene flow in peripheral populations. Evolution 51(1): 21–28.
- Garner, K.L., Chang, M.Y., Fulda, M.T., Berlin, J. ., Freed, R.E., Soo-Hoo, M.M., Revell, D.L., Ikegami, M., Flint, L. E., Flint, A.L., Kendall, B.E. 2015 Impacts of sea level rise and climate change on coastal plant species in the central California coast. PeerJ 2015(5): e958.
- Ge, X.J., Hsu, T.W., Hung, K.H., Lin, C.J., Huang, C.C., Huang, C.C., Chiang, Y.C., Chiang, T.Y. 2012 Inferring multiple refugia and phylogeographical patterns in *Pinus*

massoniana based on nucleotide sequence variation and DNA fingerprinting. PLoS ONE 7(8): e43717.

- Geng, Q., Wang, Z., Tao, J., Kimura, M.K., Liu, H., Hogetsu, T., Lian, C. 2021 Ocean currents drove genetic structure of seven dominant mangrove species along the coastlines of southern China. Front. Genet. 12: 615911.
- Guindon, S., Dufayard, J.F., Lefort, V., Anisimova, M., Hordijk, W., Gascuel, O. 2010 New algorithms and methods to estimate maximum-likelihood phylogenies: assessing the performance of PhyML 3.0. Syst. Biol. 59(3): 307–321.
- Han, C.C., Lai, C.H., Huang, C.C., Wang, I.C., Lin, H.D., Wang, W.K. 2022 Phylogeographic Structuring of the Kuroshio-Type Prawn *Macrobrachium japonicum* (Decapoda: Palaemonidae) in Taiwan and Ryukyu Islands. Diversity 14(8): 617.
- Hijmans, R., Phillips, S., Leathwick, J., Elith, J. 2024 dismo: species distribution modeling. R Package Version 1.3-16.
- Hsieh, C.-F., Shen, C.-F., Yang, K.-C. 1994 Introduction to the flora of Taiwan, 3: floristics, phytogeography, and vegetation. In: E. C. of the F. of Taiwan (Eds), *Flora of Taiwan*, 2nd ed. 2: 7–18). Department of Botany, National Taiwan University.
- Hu, L., Wang, J., Wang, X., Zhang, D., Sun, Y., Lu, T., Shi, W. 2024 Development of SSR markers and evaluation of genetic diversity of endangered Plant *Saussurea involucrata*. Biomolecules 14(8): 1010.
- Huang, B.H., Ruan, Y., Li, J.Q., Liao, P.C. 2015 Applying effective population size estimates of *Kandelia obovata* Sheue, Liu and Yong to conservation and restoration management. Forests 6(5): 1439–1453.
- Huang, C.C., Hsu, T.W. 2023 Low genetic diversity of the vulnerable Ormocarpum cochinchinense (Fabaceae: Papilionoideae) in Taiwan and implications for its conservation (abstract in English, text in Chinese). Taiwan J. Biodivers. 25(2): 37–54.
- Huang, C.C., Hung, K.H., Wang, W.K., Ho, C.W., Huang, C.L., Hsu, T.W., Osada, N., Hwang, C.C., Chiang, T.Y. 2012 Evolutionary rates of commonly used nuclear and organelle markers of *Arabidopsis relatives* (Brassicaceae). Gene **499(1)**: 194–201.
- Huelsenbeck, J.P., Ronquist, F. 2001 MrBayes: Bayesian inference of phylogenetic trees. Bioinformatics **17(8)**: 754–755.
- Ishikawa, S.I., Furukawa, A., Okuda, T., Oikawa, T. 1993 Germination requirements in *Carex kobomugi* (Sea Isle). J. Plant Res. 106(3): 245–248.
- Kamvar, Z.N., Tabima, J.F., Grunwald, N.J. 2014 Poppr: An R package for genetic analysis of populations with clonal, partially clonal, and/or sexual reproduction. PeerJ 2014(1): e281.
- Kardos, M., Armstrong, E.E., Fitzpatrick, S.W., Hauser, S., Hedrick, P.W., Miller, J. M., Tallmon, D.A., Chris Funk, W. 2021 The crucial role of genome-wide genetic variation in conservation. Proc. Natl. Acad. Sci. U.S.A. 118(48): e2104642118.
- Katoh, K., Rozewicki, J., Yamada, K.D. 2019 MAFFT online service: multiple sequence alignment, interactive sequence choice and visualization. Brief. Bioinform. 20(4): 1160– 1166.
- Kay, K.M., Whittall, J.B., Hodges, S.A. 2006 A survey of nuclear ribosomal internal transcribed spacer substitution



rates across angiosperms: An approximate molecular clock with life history effects. BMC Evol. Biol. **6**: 36.

- King, M.G., Horning, M.E., Roalson, E.H. 2009 Range persistence during the last glacial maximum: *Carex macrocephala* was not restricted to glacial refugia. Mol. Ecol. 18(20): 4256–4269.
- King, M.G., Roalson, E.H. 2008 Exploring evolutionary dynamics of nrDNA in *Carex* subgenus *Vignea* (cyperaceae). Syst. Bot. 33(3): 514–524.
- King, M.G., Roalson, E.H. 2009 Discordance between phylogenetics and coalescent-based divergence modelling: Exploring phylogeographic patterns of speciation in the *Carex macrocephala* species complex. Mol. Ecol. 18(3): 468–482.
- Ko, Y.Z., Shih, H.C., Ho, C.S., Chen, C.T., Hsu, T.W., Shiao, M.S., Chiang, Y.C. 2024 Assessment of genetic conservation units of an endangered glacial relict insular species, *Amentotaxus formosana*, based on fine-scale genetic structures of multiple fragmented mountainous populations in Taiwan. Front. Plant Sci., 15: 1–16.
- Lammi, A., Siikamäki, P., Mustajärvi, K. 1999 Genetic diversity, population size, and fitness in central and peripheral populations of a rare plant *Lychnis viscaria*. Conser. Biol. 13(5): 1069–1078.
- Leigh, J.W., Bryant, D. 2015 POPART: Full-feature software for haplotype network construction. Methods Ecol. Evol. 6(9): 1110–1116.
- Lesica, P., Allendorf, F.W. 1995 When are peripheral populations valuable for conservation? Conserv. Biol. 9(4): 753–760.
- Li, Y.L., Liu, J.X. 2018 StructureSelector: A web-based software to select and visualize the optimal number of clusters using multiple methods. Mol. Ecol. Resour. 18(1): 176–177.
- Liao, W., Hu, J., Peng, P. 2018 Burial of organic carbon in the Taiwan Strait. Journal of Geophysical Research: Oceans, 123(9): 6639–6652.
- Liu, C., Wang, J., Ko, Y. Z., Shiao, M. S., Wang, Y., Sun, J., Yuan, Q., Wang, L., Chiang, Y. C., Guo, L. 2024 Genetic diversities in wild and cultivated populations of the two closely-related medical plants species, *Tripterygium Wilfordii* and *T. Hypoglaucum* (Celastraceae). BMC Plant Biology 24(1): 195.
- M'Baya, J., Blacket, M.J., Hoffmann, A.A. 2013 Genetic structure of *Carex* species from the Australian alpine region along elevation gradients: Patterns of reproduction and gene flow. Int. J. Plant Sci. 174(2): 189–199.
- Madsen, T., Shine, R., Olsson, M., Wittzell, H. 1999 Restoration of an inbred adder population. Nature 402(6757): 34–35.
- Marshall, T.C., Slate, J., Kruuk, L. E. B., Pemberton, J.M. 1998 Statistical confidence for likelihood-based paternity inference in natural populations. Mol. Ecol. 7(5): 639–655.
- Miryeganeh, M., Takayama, K., Tateishi, Y., Kajita, T. 2014 Long-distance dispersal by sea-drifted seeds has maintained the global distribution of *Ipomoea pes-caprae* subsp. *brasiliensis* (convolvulaceae). PLoS ONE 9(4), e91836.
- Morjan, C.L., Rieseberg, L.H. 2004 How species evolve collectively: Implications of gene flow and selection for the spread of advantageous alleles. Mol. Ecol. 13(6): 1341–1356.
- Murray, M.G., Thompson, W.F. 1980 Rapid isolation of high molecular weight plant DNA. Nucleic Acids Res., 8(19): 4321–4325.

- **Nei, M.** 1987 Molecular Evolutionary Genetics. Columbia University Press, New York.
- Nikolic, N., Chevalet, C. 2014 Detecting past changes of effective population size. Evol. Appl., 7(6): 663–681.
- Ochoa-Zavala, M., Osorio-Olvera, L., Piñero, D., Núñez-Farfán, J. 2020 Inferring potential barriers to gene flow in tropical populations of *Avicennia germinans*. Aquat. Bot. 161: 103170.
- **Ohsako, T.** 2010 Clonal and spatial genetic structure within populations of a coastal plant, *Carex Kobomugi* (Cyperaceae). Am. J. Bot. **97(3)**: 458–470.
- Ohsako, T., Yamane, K. 2007 Isolation and characterization of polymorphic microsatellite loci in Asiatic sand sedge, *Carex kobomugi* Ohwi (Cyperaceae). Mol. Ecol. Notes 7(6): 1023– 1025.
- Palsbøll, P.J., Bérubé, M., Allendorf, F.W. 2007 Identification of management units using population genetic data. Trends Ecol. Evol. 22(1): 11–16.
- Peakall, R., Smouse, P.E. 2012 GenALEx 6.5: Genetic analysis in Excel. Population genetic software for teaching and research-an update. Bioinformatics 28(19): 2537–2539.
- Phillips, S.J., Anderson, R.P., Schapire, R.E. 2006 Maximum entropy modeling of species geographic distributions. Ecol. Modell. 190(3–4): 231–259.
- Pritchard, J.K., Stephens, M., Donnelly, P. 2000 Inference of population structure using multilocus genotype data. Genetics 155(2): 945–959.
- Rambaut, A. 2018 Molecular evolution, phylogenetics and epidemiology. FigTree ver. 1.4.4 software. Available online at: http://tree.bio.ed.ac.uk/software/figtree/.
- Rambaut, A., Drummond, A. J., Xie, D., Baele, G., Suchard, M. A. 2018 Posterior summarization in Bayesian phylogenetics using Tracer 1.7. Syst. Biol. 67(5): 901–904.
- Rana, S.K., Rana, H.K., Ghimire, S.K., Shrestha, K.K., Ranjitkar, S. 2017 Predicting the impact of climate change on the distribution of two threatened Himalayan medicinal plants of Liliaceae in Nepal. J. Mt. Sci. 14(3): 558–570.
- Riffe, E.C., Zinnert, J.C. 2024 Impact of invasive Carex kobomugi on the native dune community in a US mid-Atlantic coastal system. Biol. Invasions, 26(4): 1195–1208.
- Roalson, E.H., Friar, E.A. 2004 Phylogenetic relationships and biogeographic patterns in North American members of *Carex* section *Acrocystis* (Cyperaceae) using nrDNA ITS and ETS sequence data. Plant Syst. Evol., 243(3–4): 175– 187.
- Rozas, J., Ferrer-Mata, A., Sanchez-DelBarrio, J.C., Guirao-Rico, S., Librado, P., Ramos-Onsins, S.E., Sanchez-Gracia, A. 2017 DnaSP 6: DNA sequence polymorphism analysis of large data sets. Mol. Biol. Evol. 34(12): 3299–3302.
- Ruan, Y., Huang, B.H., Lai, S.J., Wan, Y.T., Li, J.Q., Huang, S., Liao, P.C. 2013 Population genetic structure, local adaptation, and conservation genetics of *Kandelia obovata*. Tree Genet. Genomes 9(4): 913–925.
- Savolainen, O., Lascoux, M., Merilä, J. 2013 Ecological genomics of local adaptation. Nat. Rev. Genet. 14(11): 807– 820.
- Savolainen, O., Pyhäjärvi, T., Knürr, T. 2007 Gene flow and local adaptation in trees. Annu.l Rev. Ecol. Evol. Syst. 38(1): 595–619.
- Schlotterer, C. 2000 Evolutionary dynamics of microsatellite DNA. Chromosoma **109(6)**: 365–371.



- Schönswetter, P., Elven, R., Brochmann, C. 2008 Trans-Atlantic dispersal and large-scale lack of genetic structure in the circumpolar, arctic-alpine sedge *Carex bigelowii* s. l. (Cyperaceae). Am. J. Bot. **95(8)**: 1006–1014.
- Senni, K., Fuji, N., Takhashi, H. 2005 Intraspecific chloroplast DNA variations of the alpine plants in Japan. Acta Phytotaxon. Geobot. 56(3): 265–275.
- Sexton, J.P., Strauss, S.Y., Rice, K.J. 2011 Gene flow increases fitness at the warm edge of a species' range. Proc. Natl. Acad. Sci. U.S.A. 108(28): 11704–11709.
- Shaw, J., Lickey, E. B., Beck, J. T., Farmer, S. B., Liu, W., Miller, J., Siripun, K. C., Winder, C. T., Schilling, E. E., Small, R. L. 2005 The tortoise and the hare II: Relative utility of 21 noncoding chloroplast DNA sequences for phylogenetic analysis. Am. J. Bot. 92(1): 142–166.
- Suárez, D., Arribas, P., Jiménez-García, E., Emerson, B.C. 2022 Dispersal ability and its consequences for population genetic differentiation and diversification. Proc. R. Soc. B Biol. Sci. 289(1975): 289.
- Takayama, K., Kajita, T., Murata, J., Tateishi, Y. 2006 Phylogeography and genetic structure of *Hibiscus tiliaceus*-Speciation of a pantropical plant with sea-drifted seeds. Mol. Ecol. 15(10): 2871–2881.
- Takayama, K., Tateishi, Y., Murata, J., Kajita, T. 2008 Gene flow and population subdivision in a pantropical plant with sea-drifted seeds *Hibiscus tiliaceus* and its allied species: Evidence from microsatellite analyses. Mol. Ecol. 17(11): 2730–2742
- Wahlsteen, E. 2021 SSR markers distinguish critically endangered *Acer campestre* populations from cryptic invading gene pools. Willdenowia 51(1): 115–125.
- Wang, J.C., Chen, H.H., Hsu, T.W., Hung, K.H., Huang, C.C. 2024 A taxonomic revision of the genus *Angelica* (Apiaceae) in Taiwan with a new species *A. aliensis*. Bot. Stud. 65(1): 3.
- Wee, A.K.S., Takayama, K., Asakawa, T., Thompson, B., Onrizal, Sungkaew, S., Tung, N.X., Nazre, M., Soe, K.K., Tan, H.T. W., Watano, Y., Baba, S., Kajita, T., Webb, E. L. 2014 Oceanic currents, not land masses, maintain the genetic structure of the mangrove *Rhizophora mucronata* Lam. (Rhizophoraceae) in Southeast Asia. J. Biogeogr. 41(5): 954–964.
- WFO 2024 Carex kobomugi Ohwi. Published on the Internet;Http://Www.Worldfloraonline.Org/Taxon/Wfo-0000347956. Accessed on: 02 Aug 2024.
- Willyard, A., Syring, J., Gernandt, D. S., Liston, A., Cronn, R. 2007. Fossil calibration of molecular divergence infers a moderate mutation rate and recent radiations for Pinus. Mol. Biol. Evol. 24(1): 90–101.

- Wilson, G.A., Rannala, B. 2003 Bayesian inference of recent migration rates using multilocus genotypes. Genetics, 163(3): 1177–1191.
- Wolfe, K.H., Li, W.H., Sharp, P.M. 1987 Rates of nucleotide substitution vary greatly among plant mitochondrial, chloroplast, and nuclear DNAs. Proc. Natl. Acad. Sci. U.S.A. 84(24): 9054–9058.
- Wootton, L.S., Halsey, S.D., Bevaart, K., McGough, A., Ondreicka, J., Patel, P. 2005 When invasive species have benefits as well as costs: Managing *Carex kobomugi* (Asiatic sand sedge) in New Jersey's coastal dunes. Biol. Invasions 7(6): 1017–1027.
- Yamamoto, T., Tsuda, Y., Takayama, K., Nagashima, R., Tateishi, Y., Kajita, T. 2019 The presence of a cryptic barrier in the West Pacific Ocean suggests the effect of glacial climate changes on a widespread sea-dispersed plant, *Vigna marina* (Fabaceae). Ecol. Evol. 9(15): 8429–8440.
- Yan, H., Qi, H., Li, Y., Wu, Y., Wang, Y., Chen, J., Yu, J. 2022 Assessment of the genetic relationship and population structure in oil-Tea *Camellia* species using simple sequence repeat (SSR) markers. Genes, 13(11): 2162.
- Yang, H., Lu, Q., Wu, B., Zhang, J. 2012 Seed dispersal of East Asian coastal dune plants via seawater - short and long distance dispersal. Flora Morphol. Distrib. Funct. Ecol. Plants 207(10): 701–706.
- Yano, O., Ikeda, H., Hoshino, T. 2010 Phylogeography of the Japanese common sedge, *Carex conica* complex (Cyperaceae), based on chloroplast DNA sequence data and chromosomal variation. Am. J. Bot., **97(8)**: 1365–1376.
- Yu, G. 2020 Using ggtree to visualize sata on tree-like structures. *Current Protocols in* Bioinforma. 69(1): e96. h
- Zhang, X., Liu, Y. H., Wang, Y.H., Shen, S.K. 2020 Genetic diversity and population structure of *Rhododendron rex* subsp. *rex* inferred from microsatellite markers and chloroplast DNA sequences. Plants 9(3): 338.
- Zhang, X., Yang, L., Liu, Y.H., Zhou, X.L., Zhang, L.Q., Wang, Y.H., Shen, S.K. 2021 Genetic diversity, genetic structure, and demographic history of *Cinnamomum chago*, a plant species with extremely small populations in China. Glob. Ecol. Conserv. **31**, e01808.
- Zimmerman, S.J., Aldridge, C.L., Oyler-Mccance, S.J. 2020 An empirical comparison of population genetic analyses using microsatellite and SNP data for a species of conservation concern. BMC Genomics **21(1)**: 1–16.

Supplementary materials are available from Journal Website