



NOTE

Lectotypification, morpho-anatomical traits and initial chemical analysis of *Psychotria condorensis* Pierre ex Pitard (Rubiaceae, *Psychotria*): A study on an endemic species from Con Dao National Park, Ba Ria-Vung Tau Province, Vietnam

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(Manuscript received 30 April 2024; Accepted 30 August 2024; Online published 6 September 2024)

ABSTRACT: *Psychotria condorensis* Pierre ex Pitard is an endemic species described from Con Dao National Park, Ba Ria-Vung Tau province, Vietnam. This study provides the lectotype of this species and describes its micromorphological along with phytochemical characteristics. These results are used as a monograph in the identification and standardization of medicinal materials, conservation assessment of this species, and contributes information for advancing further research on conservation and its biological activities.

KEY WORDS: Anatomy, morphology, plant diversity, plant taxonomy, lectotype.

INTRODUCTION

The genus *Psychotria* Linné is one of the largest angiosperm genera in the world, estimated to include about 2000 species globally (Dwyer, 1980; Davis, 2001; Taylor *et al.*, 2007). *Psychotria* species are extensively distributed in tropical and subtropical regions, including Africa and Madagascar. They exhibit well-described morphological characteristics such as being shrub, small tree, or climber, with opposite leaves, entire or bilobed stipules that are deciduous, and inflorescences in the form of cymes, racemes, or thyrses, featuring sessile or pedicellate entomophilous or distylous flowers. Their fruits are shiny with two plano-convex pyrenes, and seeds can possess ruminant or abundant albumin (Hamilton 1989, Nepokroeff *et al.*, 1999, Andersson, 2002, Chen and Taylor, 2011).

Pharmacological studies have shown that the *Psychotria* genus has a variety of biological activities, including antimicrobial, antiviral, analgesic, hypoglycemic, and strong-cancer cell-killing effects (Benevides *et al.*, 2005; Pimenta, *et al.*, 2011; Tran *et al.*, 2024). The chemical composition studies have shown that the *Psychotria* genus contains group of compounds such as alkaloid (*Psychotria malayana* Jack, *Psychotria suterella* Müll. Arg. etc.), flavonoids (*Psychotria yunnanensis* Hutch.), terpenoid (*Psychotria rubra* (Lour.) Poit., *Psychotria yunnanensis* Hutch., etc.), steroids (*Psychotria hainanensis* H.L.Li, etc.), coumarin (*Psychotria stachyoides* Benth.), tannins, organic acids, amino acids, reducing sugars, polysaccharides, fats,

sterols, carotenoids, phenolic compounds, and saponins (Tran, 2018).

In Vietnam, there are estimated to be 26 species within this genus, along with one variety (Pham, 2000; Tran, 2005; Nguyen *et al.*, 2023b), including 9 known species with medicinal properties to treat respiratory and epidemic-related diseases (Vo, 2012). During our botanical survey in Con Dao National Park, Ba Ria-Vung Tau Province, Vietnam, several unknown *Psychotria* specimens were collected. After carefully examining relevant taxonomical literatures (Pitard, 1924; Pham, 2000; Tran, 2005; Chen and Taylor, 2011) and comparing morphological characters in available herbaria and digital herbarium materials of *Psychotria* specimens e.g. P, K, JSTOR Global Plants, the species matches to *Psychotria condorensis* Pierre ex Pitard (1924). This species was first described in "Flore générale de l'Indo-chine" by Pitard (1924), and it was mentioned in the Flora of Vietnam (Pham, 2000; Nguyen, 2005). However, during our field survey, we noted some distinct characteristics that were not mentioned in the original description, especially when examining specimens in various museums where the lectotype of the species has not been recorded. Furthermore, current anatomical research on this species as well as this genus is also very limited, except for common species such as *Psychotria rubra* (Lour.) Poir. (synonym of *Psychotria asiatica* L.) (Bui and Tran, 2013), *Psychotria adenophylla* Wall. (Nguyen *et al.*, 2023a), e.g. Based on the aforementioned reasons, this study is conducted with the purpose of providing relative information on designing a lectotype specimen for the



species in accordance with the Shenzhen Code Art. Art. 9.3, 9.4, 9.7, 9.12 (Turland *et al.*, 2018). Additionally, we provide an amended description of this species based on our recently collected materials, a color photograph, preliminary conversation status, and some notes of this species, the morphological characteristics and microanatomy are being examined through optical microscopy, and initial investigations into the active ingredients are being conducted to reinforce taxonomic studies as well as to explore the potential medicinal properties of this genus in the future.

MATERIAL AND METHODS

Macro-morphology. The studied materials were collected from Con Dao National Park, Ba Ria – Vung Tau province, and preserved in ethanol 70°. All photographs were captured using Canon EOS 750D mounted with Canon EF-S 60 mm f/2.8 Macro USM lens. The morphological terminology is based on Beentje (2012). Specimens and literature relevant to *Psychotria* species from the Indo-china and neighboring countries were reviewed in various herbaria using high-resolution images from Global Plants on JSTOR, available at <https://plants.jstor.org/>, and through the Global Biodiversity Information Facility (GBIF), accessible at <https://www.gbif.org>. The morphological description's nomenclature is in line with Beentje (2012). The evaluation of conversations was conducted in accordance with the criteria set forth by the International Union for Conservation of Nature (IUCN, 2012) criteria.

Micro-morphology. The materials (including young stems, mature stems, and leaves) were manually sliced into thin sections using a razor blade. The transverse sections were clarified using chloral hydrate (8%) and acidified with acetic acid (1%). These sections were then stained with Carmine (10%) in 30 minutes and Methylene Blue (1%) in 15 seconds. The excess reagents were gradually removed using distilled water on three times, following the method described by Tran (1981). The sections were placed on slides in glycerin (10%) and covered with a slip. Finally, the prepared slides were examined under a light microscope at 10X and 40X magnifications using a Nikon Eclipse E100 (Japan).

Power characteristics. According to the method of Nguyen (2007) and Nguyen (2020). Leaves were collected, washed and dried under 50°C until constant mass, then ground into a fine powder, which was then sifted using the appropriate mesh size. The leaf flour characteristics were placed on slides in water and were observed under microscope (Nikon Eclipse E100, Japan).

Preliminary phytochemical screening. The leaf powder samples were extracted using different solvents have varying degrees of polarity, and preliminary qualitative chemical analysis was conducted based on several characteristic chemical reactions Nguyen (2007),

Nguyen (2020). Different phytochemical components, such as alkaloids, flavonoids, tanins, saponins, carbohydrates, coumarins, amino acids, fats, cardiac glycosides, proanthocyanidins, anthocyanosides, polyuronides were examined for their qualitative characteristics following established procedures.

RESULTS

Anatomical characteristics

Midrib leaf-anatomy. In a sectional view, the midrib exhibits a convex shape on its lower sides (Fig. 1). The upper and lower leaf epidermis consists of a single layer and is externally shielded by a thin cuticle layer (line 1). The upper epidermis cells (line 2) are slightly larger compared to those in the lower epidermis (line 10). The lower epidermis cells, arranged adjacent to one another, take on a triangular shape of varying sizes (line 10). The outermost wall undergoes thickening and is concealed by either invisible or attenuated hair in a cross-section of the midrib (line 11). The lower collenchyma comprises 7–9 layers of isodimetric cells with thickened walls (line 9). The lower parenchyma, with 13–15 layers, features circular or sub-circular cells of assorted sizes, thin walls, and a random arrangement (line 8). The central bundle adopts a V-shaped structure, encompassing downward phloem with randomly arranged dark-pink cells (line 6), while the upward phloem's xylem is constituted by large, aligned hollow cells with thickened, lignified, and blue-colored walls (line 7), and peripheral vascular bundle sheath consists of lignified cells, blue-colored walls (line 5). The upper epidermis with one layer, upper collenchyma with 5–6 layers (line 3), and upper parenchyma (line 4) with 5–6 layers are closely mirror the structure observed in the lower layers

Leaf blade-anatomy. In the leaf blade (Fig. 2A), the upper epidermis (line 1) consists of one layer with triangular cells that are large-sized, thin-walled, and externally shielded by a thin cuticle layer. The hypodermis, located beneath the upper epidermis, is composed of polygonal cells in 1–2 layers, which are large-sized and thin-walled (line 2). The mesophyll is dorsiventral and comprises one layer of compactly arranged palisade parenchyma cells (line 3) and up to ten layers of spongy parenchyma cells (line 4), which are loosely arranged with large airspaces (line 8), moreover, the starch are found in the palisade parenchyma (line 7). The lower epidermis has small, papillae cells in one layer, which are thin-walled (line 5). The surface of the lower epidermis shows sporadic non-glandular trichomes (line 9). The stomata are confined to the lower surface and are located above the level as the adjacent epidermal cells (Fig. 4B-C, line 6).

Young stem-anatomy. The transverse section of the young stem was elliptical to sub-circular in shape (Fig. 3A), covered by a single epidermal layer composed of

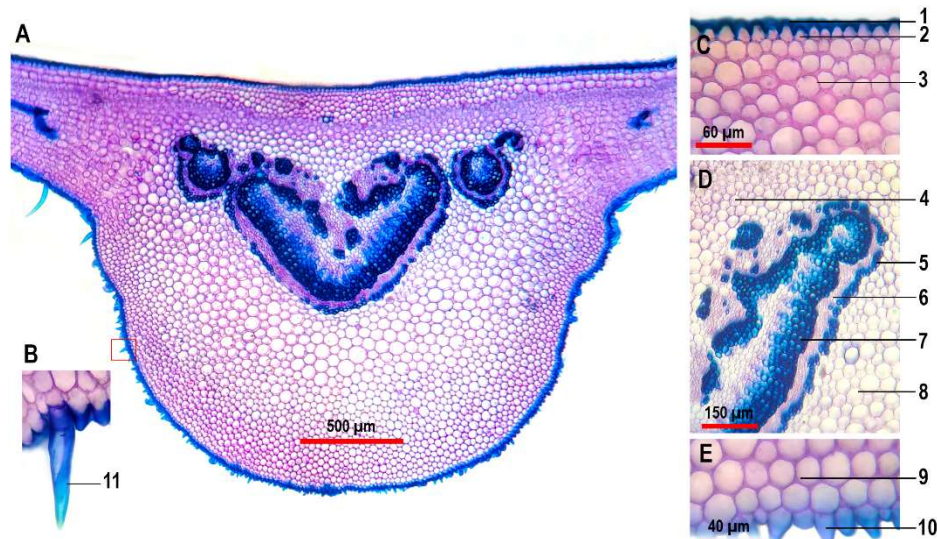


Fig. 1. Microscopic characteristics of midrib leaf cross-sections of *P. condorensis*. **A.** Midrib cross-section. **B-E.** Enlarging the internal structures of leaf midrib. 1. Cuticle. 2. Upper epidermis. 3. Upper collenchyma. 4. Upper parenchyma. 5. Bundle sheath. 6. Phloem. 7. Xylem. 8. Lower parenchyma. 9. Lower collenchyma. 10. Lower epidermis. 11. Trichomes.

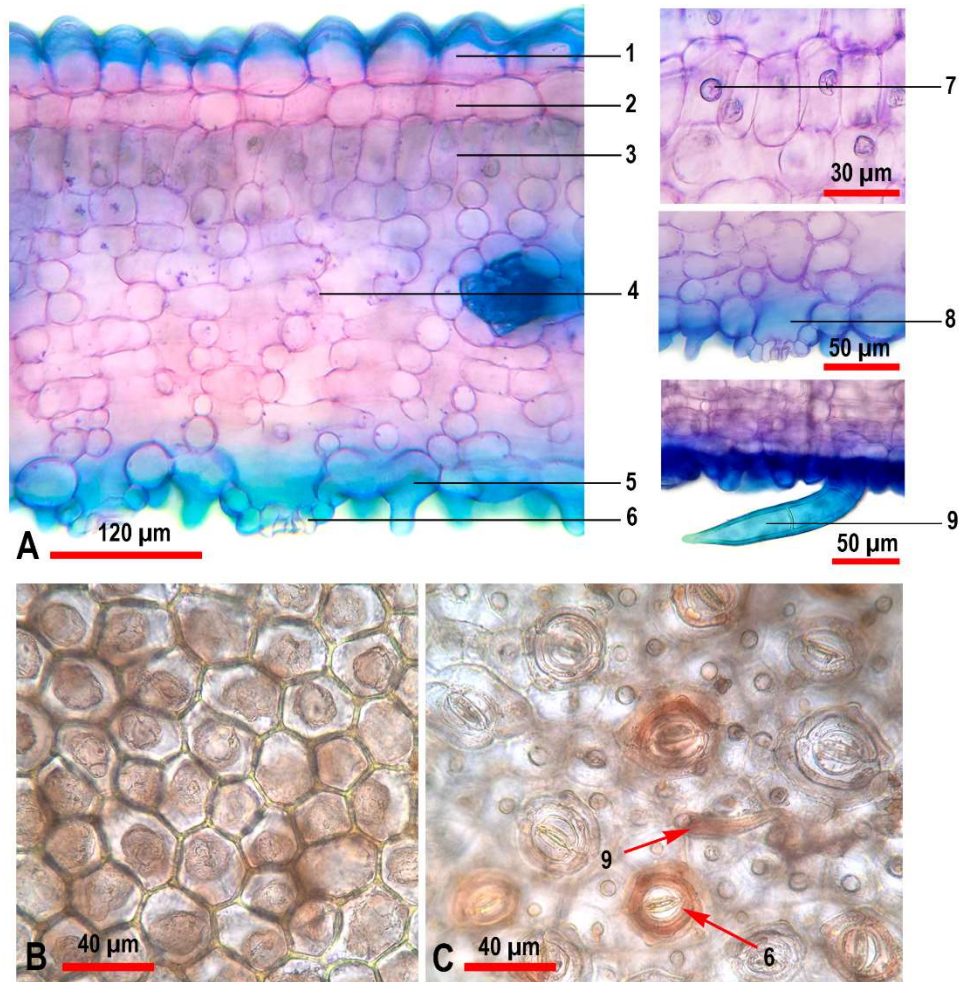


Fig. 2. Microscopic characteristics of leaf blade cross-sections of *P. condorensis*. **A.** Blade cross-sections and enlarging the internal structures of leaf blade. **B.** The adaxial leaf surfaces. **C.** The abaxial leaf surfaces. 1. Upper epidermis. 2. Hypodermis. 3. Palisade parenchyma. 4. Spongy parenchyma. 5. Lower epidermis. 6. Stomata. 7. Starch. 8. Sub-stomatal chamber. 9. Trichomes.

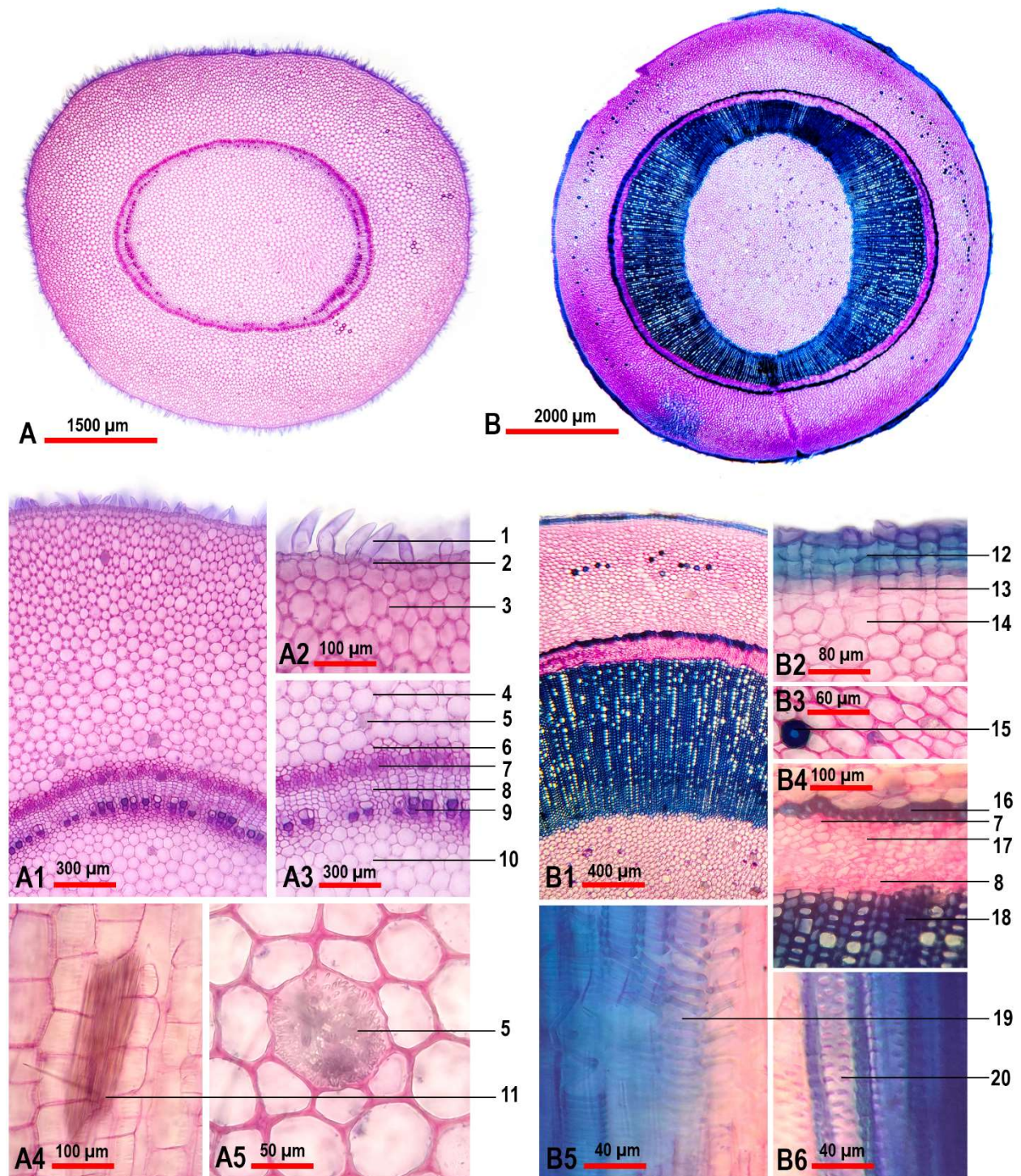


Fig. 3. Microscopic characteristics of stems cross-sections of *P. condorensis*. **A, A1-A5.** Stem cross-sections and enlarging the internal structures of young stems. **B, B1-B6.** Stem cross-sections and enlarging the internal structures of mature stems. 1. Non-glandular trichomes. 2. Epidermic. 3. Collenchyma. 4. Parenchyma. 5. Needle-shaped crystals cross-section. 6. Sclerenchyma. 7. Primary phloem. 8. Vascular bundle. 9. primary xylem. 10. Pith parenchyma. 11. Needle-shaped crystals longitudinal-section. 12. Phellem. 13. Cork cambium bundle. 14. Phelloderm. 15. Fibers. 16. Sclerenchyma cells have thick walls that undergo lignification to form fibers. 17. Secondary phloem. 18. Secondary xylem. 19. Spiral vessel. 20. scalariform vessel.



small, adjacent, polygonal-shaped cells (Fig. 3A2, line 2) with a thick outermost wall containing non-glandular trichomes (Fig. 3A2, line 1). The collenchyma consisted of adjacent, polygonal-shaped cells in 10-12 layers, with thickened walls at the corners (Fig. 3A1-A2, line 3). Following the collenchyma, the cortical parenchyma composed multiple layers with randomly organized circle-shaped cells of varying sizes and thin walls (Fig. 3A1, line 4). The sclerenchyma included small, polygonal-shaped cells in a single layer, covering and distinguishing between the cortical parenchyma and the vascular bundle (Fig. 3A3, line 6). The vascular bundle comprised primary phloem and primary xylem arranged in an annular pattern. The externally located primary phloem contained randomly arranged polygonal cells, smaller in size and darker in color than parenchyma cells (Fig. 3A3, line 7). Internally, the primary xylem included vessels arranged radially (Fig. 3A3, line 9). The vascular cambium, situated between the primary phloem and primary xylem, consisted of rectangular cells in four layers, thin-walled, smaller in size, and arranged longitudinally (Fig. 3A3, line 8). The pith parenchyma was composed of sub-circular cells that were thin-walled, large, and arranged randomly (Fig. 3A3, line 10). In addition, the needle-shaped crystals were found scattered in parenchyma, collenchyma of young stem (Fig. 3A4-A5, line 5&11).

Mature stem-anatomy. In conjunction with the growth of cork cambium and vascular cambium, the stem developed in terms of shape and size. The structure of the transverse section of the mature stem (Fig. 3B) is similar to that of the young stem but is distinguished by the presence of periderm (Fig. 3B2), fiber sclerenchyma, and a second vascular bundle (Fig. 3B1). The cork cambium (Fig. 3B2, line 13) actively produces phellem (Fig. 3B2, line 12) on the outer side and phelloderm on the inner side (Fig. 3B2, line 14). The phellem consists of rectangular cells in two layers, adjacent and arranged to absorb a blue color; phelloderm is created beneath the cork cambium, including rectangular cells in a single layer with a thin wall and a lighter color than collenchyma and parenchyma. The sclerenchyma cells have thick walls that undergo lignification to form fibers, covering the vascular bundle (Fig. 3B4, line 16). The function of the vascular bundle (Fig. 3B3, line 8) produces secondary phloem (Fig. 3B3, line 17) and secondary xylem (Fig. 3B3, line 18). The secondary phloem externally, composed of polygonal cells with thin, wrinkled walls. The secondary xylem inward consists of large xylem vessels (spiral vessels and scalariform vessels) randomly arranged in a radial pattern. Distinguishing between the primary phloem (Fig. 3B4, line 7) and primary xylem is challenging because they are flattened by the secondary bundle. Furthermore, fibers are found in the cortical parenchyma of the mature stem (Fig. 3B3, line 15). In the longitudinal section of the secondary xylem has two types,

including spiral vessel (Fig. 3B5, line 19) and scalariform vessel (Fig. 3B6, line 20).

Leaf powder. The brownish powdered leaf, having a gentle fragrance, had several microscopic characteristics, including unicellular and multicellular non-glandular trichomes (Fig. 4H & 4I). Fragments of the upper epidermis (Fig. 4A) and lower epidermis containing small stomata with two smaller guard cells were found under the microscope (Fig. 4B). Spiral (Fig. 4F) and scalariform vessel pieces (Fig. 4G) were found in the powdered material. Fragmented collenchyma (Fig. 4C), parenchyma (Fig. 4D), fiber (Fig. 4E) rarely appeared, and the circular starch granules were approximately 10-20 µm in diameter (Fig. 4K). The needle-shaped crystals were scattered or gathered in clusters that were approximately 150-200 µm long (Fig. 4J).

Preliminary analysis of the chemical composition of *Psychotria condorensis* Pierre ex Pitard leaf powder

The result of preliminary phytochemical analysis of leaf extract of *Psychotria condorensis*, including it shows the presence of flavonoids, tannins, saponins, carbohydrates, coumarins, amino acids, and negative test for alkaloids, fats, cardiac glycosides, proanthocyanidins, anthocyanosides, polyuronides (Table 1).

Table 1. Phytochemical screening of *Psychotria condorensis* leaves.

| Chemical constituents | The test | Result | Conclusion |
|---------------------------|--------------------------------------|--------|------------|
| Alkaloids | Wagner's test | (-) | Absent |
| | Dragendroff test | (-) | |
| Flavonoids | Cyanidin test | (+) | Present |
| | FeCl ₃ 5% test | (+) | |
| Tannins | FeCl ₃ 5% test | (+) | Present |
| | Gelatin 1% test | (-) | |
| | Lead acetate 10% test | (+) | |
| Fats | Stain test | (-) | Absent |
| Saponins | Foam test | (+) | Present |
| Carbohydrates | Fehling's test | (+) | Present |
| Cardiac glycosides | Keller – Kiliani's test | (-) | Absent |
| Coumarins | Flourescences test | (+) | Present |
| Amino acids | Na ₂ CO ₃ test | (+) | Present |
| Proanthocyanidins | HCl 10% test | (-) | Absent |
| Anthocyanosides | HCl 10% + NaOH | (-) | Absent |
| | 10% test | (-) | |
| Polyuronides | Ethanol 95° test | (-) | Absent |

TAXONOMIC TREATMENT

Psychotria condorensis Pierre ex Pitard, Fl. Indo-Chine 3: 351 (1924); Pham, H.H. *P. condorensis*, An Ill. Fl. of Viet. III: 195 (2000); Tran, N.N. *P. condorensis*, in Nguyen, T.B. eds., Checkl. Pl. Spec. Vietn. 3: 141 (2005).

Type: VIETNAM. Ba Ria-Vung Tau Province, Con Dao National Park, *Cochinchiaad uisulam Condor*. Sep. 1876, Dr. Harmand No. 1932 (lectotype P [P04020420 –

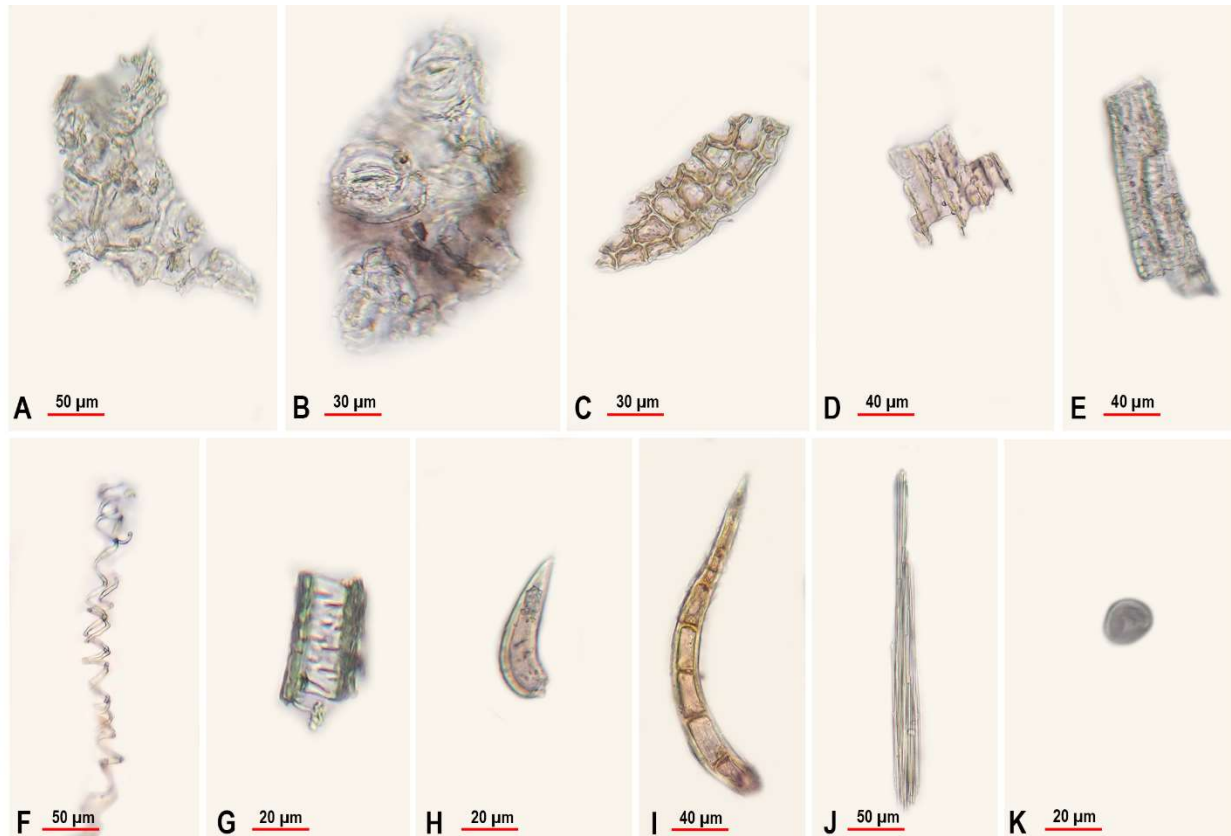


Fig. 4. Microscopic features of leaf powder. **A.** piece of upper epidermis. **B.** piece of lower epidermis. **C.** Fragment of collenchyma. **D.** fragment of parenchyma. **E.** Fiber. **F.** Fragment of spiral vessels. **G.** Fragment of scalariform vessel. **H.** unicellular non-glandular trichomes. **I.** multicellular non-glandular trichomes.

<http://coldb.mnhn.fr/catalognumber/mnhn/p/p04020420>], **Fig. S1**; isoelectotype: P04020421; K000777254 - <https://data.kew.org/records/occurrences/1a2e55f9-6880-4784-9f7c-477fac50c353>), **designated here**).

Description. **Shrubs**, 1–3.5 m tall. **Twigs** puberulent when young, glabrous when mature, reddish brown to gray *in vivo*, dark brown *in sicco*, stems sub-cylindrical to cylindrical, internodes 1.0–5.2 cm long. **Stipules** ovate-triangular, 5–6 mm long, green to yellowish-green when young, dark brown when dry, glabrous on both sides, apex acute to obtuse, base large oval, margin entire, caducous. **Leaves** simple, opposite-decussate, petiolate; lamina elliptic-oblong, lanceolate-oblong to obovate, 10.5–28.5 × 5.5–9.5 cm, upper surface green and lower surface light green *in vivo*, turning blackish-brown when dried, glabrous on adaxial surfaces, glabrous to very sparsely hairs on abaxial surfaces, base cuneate, apex acuminate to acute, margin entire, venation mixed brochidodromous, midrib prominent on abaxial surface, glabrous to puberulent; secondary veins 15–20 pairs, without domatia; petioles 1.0–2.2 cm long, glabrous to puberulent. **Inflorescence** terminal in new branches, cyme, erect paniculate; peduncle 2–5 cm long, puberulent, 3–4 times dichotomous, primary branches 1.2–2.1 cm long, puberulent, secondary branches 5–10 mm long,

puberulent, green to dark green *in vivo*; bracts short ovate-triangular, puberulent on outer surface, glabrous on inner surface, apex obtuse, caducous. **Flowers** pedicellate; pedicel 1–3.5 mm long, glabrous to puberulent, bracteoles short-ovate, short, caducous, 1.4–1.8 mm long, puberulent on outer surface, glabrous on inner surface, apex obtuse, margin entire, caducous. **Calyx** white, cupuliform, glabrous, tube 1.4–1.9 mm long, lobes null, truncated at the top, on board barely sinuous. **Corolla** white, tube 6–8 mm long, glabrous; lobes 5–6 merous, ovate-triangular, apex acute, 3–4.5 mm long, thick and recurved at the apex, puberulent in tip outer surface, glabrous on inner surface; densely villous from throat to middle tube inside. **Stamens** 5–6, inserted, filaments very short, glabrous, erect; anthers oblong-elliptic, 1.5–1.8 mm long, apex obtuse, dorsifixed. **Ovary** 2-locular, ovule 1 in each locule; style 7–8 mm long, glabrous to puberulent; stigma 2-lobed, ca. 1 mm long. **Fruits** fleshy, globoid to ovoid, 8–11 mm long, 6–8 mm in diam., green when young, orange when ripe, glabrous, apex with persistent calyx lobes; pyrene 2, plan ventral, convex dorsal. **Seeds** semi-ellipsoid to semi-ovoid, 3–5 mm long, 3–5 mm wide, blackish brown when dry, albumin ruminant (Fig. 5).

Distribution and habitat. *Psychotria condorensis* was so far only found from its type locality, in tropical

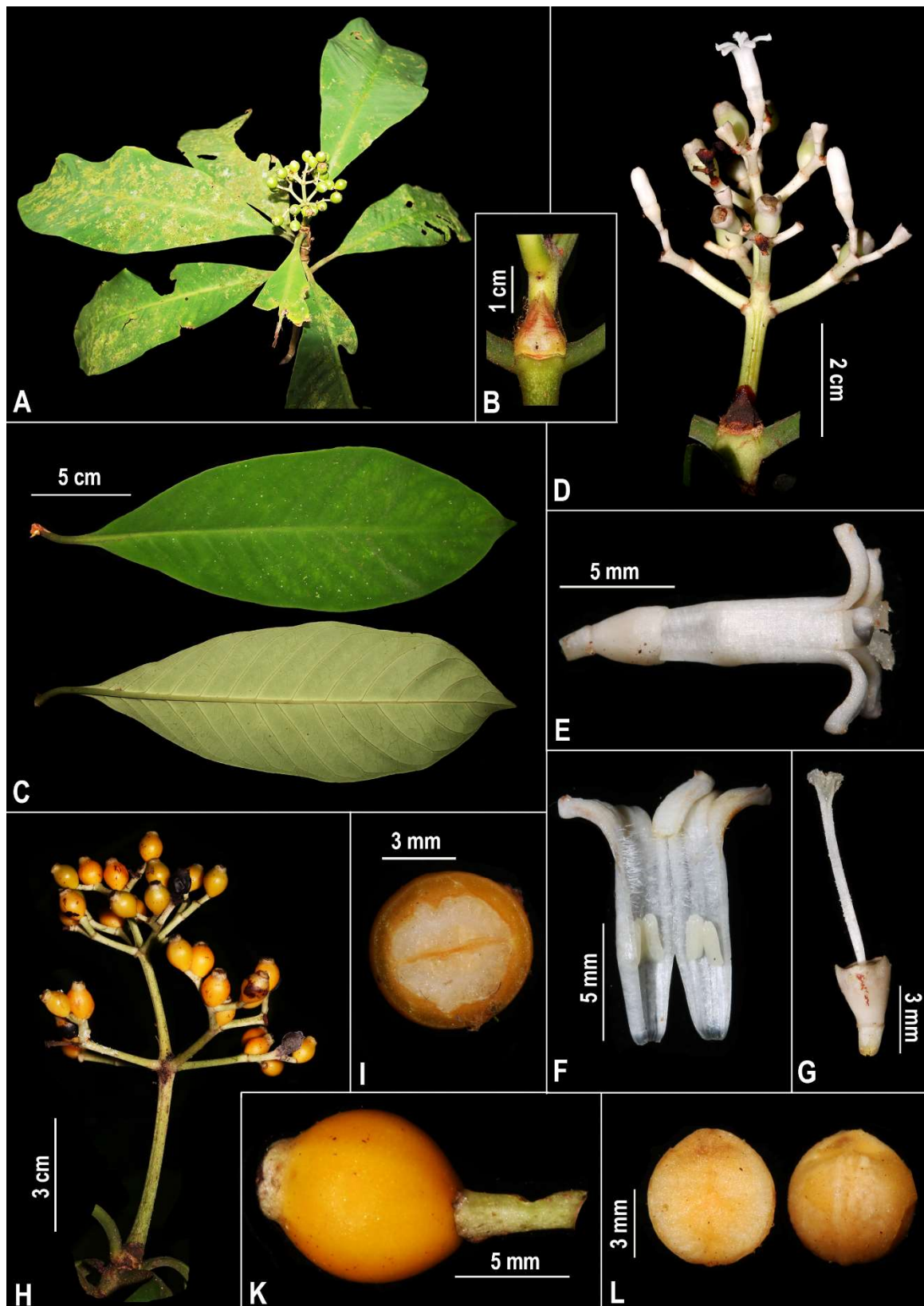


Fig 5. *Psychotria condorensis* Pierre ex Pitard. **A.** Branch brings young fruitlets. **B.** Stipule. **C.** Leaves (Adaxial surface and abaxial surface). **D.** Inflorescence. **E.** Flower. **F.** Corolla cross-section whows stamens. **G.** Calyx, style and stigma. **H.** Fruitlet. **I.** Fruit cross-section shows 2-pyrenes. **K.** Mature fruit. **L.** Pyrenes (Photos by Nguyen Quoc Bao).



evergreen forest in Con Dao National Park, Ba Ria – Vung Tau province, at elevation 250 m to 550 m a.l.s.

Phenology. Flowering and fruiting from January to May.

Vernacular name. Lầu Côn sơn.

Preliminary conservation status. *Psychotria condorensis* is the endemic species in Con Dao National Park, Vietnam. From our field observations, the estimated number of mature individuals is approximately under 50. The species has an extent of occurrence (EOO) of less than 1 km² and an area of occupancy (AOO) of less than 8 km². The habitat of *P. condorensis* is within the protected area of the Nation Park, which is well preserved and stable. According to the IUCN Red List criteria D, its status is Critically Endangered (CR) (IUCN, 2019).

Specimen examined. VIETNAM, Ba Ria-Vung Tau province, Con Dao National Park, elevation 255 m, March 2022, *T. B. Quyen et al.*, Dang 509 (VNM00070357); Ba Ria-Vung Tau province, Con Dao National Park, Mt. Thanh Gia, elevation 555 m, March 2024, *N. Q. Bao.*, QB 115 (VNM00070358); VIETNAM, Ba Ria-Vung Tau province, Con Dao National Park, “Expédition du Dr Harmand en Indo-Chine 1875-77, Iles de Poulo-Condor (Cochinchine française)”, September 1876, Harmand N°847 det. C.J.Pitard (P04020418 - <https://science.mnhn.fr/institution/mnhn/collection/p/item/p04020418?listCount=4&listIndex=1>, P04020419-<http://coldb.mnhn.fr/catalognumber/mnhn/p/p04020419>).

Taxonomic notes. This entity was originally named as *Grumilea condorensis* (unpublished mss.) by J.P.L. Pierre. Subsequently, Pitard (1924) gave it a formal name based on Harmand’s specimens from Condor (a group of island in Southern Vietnam which is now called Con Dao in Vietnamese) as *Psychotria condorensis* and described it in in Flore Générale de l’Indo-chine, clearly stating the specimen, ‘COCHINCHINE: Condor (Harmand)’. This voucher corresponds with two specimens from the collections of L. Pierre at the Muséum national d’Histoire naturelle (P), viz., Cochinchina, island of Condor, September 1876, Dr. Harmand No. 1932 (P04020420, and P04020421). Among them, two sheets P04020420 and P04020421 are represents complete specimens with hand written latin description, while P04020420 also contains a small line drawing of pyrenes i.e. seeds (see fig. 1). The label on P04020420, can be read as: “N°. 1932 *Grumilea condorensis* Pierre; *Psychotria* auct.; Flores 5–6 meri; Costulae 16–20 utrinque. Drupa lobis vel ecostata. Pyrena plano convexus dorso praeter basin compell ovata obtuse carinata, finata, ad ventrem plana. Semen plano convexum, albumine valde commixtum; Hab Cochinchina ad insulam Condor; Stylus 8 ½ mm longa, lobis stigmatibus breviter 1 ½ cm lectus barbatis vel glabris; Coll. Dr. Harmand; 9/1876”. On the sheet P04020421, similar handwritten description is also present. On the opposite side, the label “HERB. L. PIERRE” following text is written: “N°. 1932 *Grumilea condorensis* Pierre; *Psychotria kleusa* (?) Hook.f. ≠ Pierre; *Uragoga seusa* H.Br (?); Hab. Crescitui um. Condor.; 9/1876; Coll. Dr Harmand”. There are third and fourth specimen (P04020418, P04020419) with similar label, however, it mentions *Psychotria* 847 and there is no mentioned of

number 1932, neither does it contain and latin annotations. We can’t confirm that this is a part of the same collection No. 1932, hence, we treat it as a distinct collection which was also studied by Pitard (1924) while he was describing this species.

At Kew Herbarium (K), there is a fifth specimen K000777254 which also originates from “HERB. L. PIERRE” and contains further information as: “N°. 1932 *Grumilea condorensis* Pierre; *Psychotria condorensis* Pierre; Hab. Crescitui um. Condor.; 9/1876; Coll. Dr Harmand”. Given this situation, based on Art. 9.3, 9.4, 9.12 of the International Code of Nomenclature for algae, fungi, and plants (Shenzhen Code) (Turland *et al.* 2018), we have chosen the sheet from Harmand’s collection in the P herbarium [P04020420] as the lectotype, as it represents a complete specimen, long latin description and line drawing of the seed. While rest of the specimens belonging to the collection No. 1932, viz. P04020421 and K000777254 are treated as Isolectotype. *Psychotria* 847 (P04020418) is treated as lectoparatype, and P04020419 as Isolectoparatype as per Art. 9.7 of the Shenzhen Code (Turland *et al.* 2018).

DISCUSSIONS

Psychotria condorensis is one of the species with a narrow distribution, only found outside of Con Dao, Vietnam, at altitudes ranging from 250 to 550 meters. With a low number of mature individuals, it is considered endemic and in need of conservation in this area. Morphologically, it aligns with Pitard’s initial description, but this study provides additional anatomical details such as: young twigs puberulent, mature twigs glabrous, abaxial leaves blade sparsely hairs, petioles glabrous to puberulent, cyme inflorescences, white flowers long-styled morphs with long corolla tube, style glabrous to puberulent, smooth characteristic orange ripe fruits.

The anatomical characteristics of *Psychotria condorensis* adhere to classical dicotyledonous anatomy, showcasing dorsiventral leaf organization, a lignified stem exhibiting the conventional sequence of periderm, secondary phloem, vascular cambium, secondary xylem replete with vessel elements, and a central pith within the stem. The anatomical features of the leaf blade were initially described, with an average thickness of approximately 400–450 micrometers. The leaf anatomy of this species is typical of a shrub, often found growing under the canopy of forests. The palisade layer includes one player, while the spongy layer is thicker and with prominent air spaces. In this study, the larger upper epidermic covered with cuticle which are reported present on the adaxial surface of *P. leiocarpa*, *P. glaziovii*, *P. hoffmannseggiana* (Moraes *et al.*, 2011). The existence or lack of cuticular ornamentation has previously been documented in *Psychotria* species that reside in moist, shaded areas. (Quinteiro *et al.*, 2006). The hypodermis



under the upper epidermic, this observation was first documented, the hypodermis is one of the characteristic structures in the species, serving a storage function. According to previous anatomical reports, the hypodermis absent in other species, such as: *P. fractistipula* L.B.Sm., R.M.Klein & Delprete (de Oliveira *et al.*, 2020), *P. deflexa* DC., *P. leiocarpa* Cham. &, *P. racemosa* Rich. (Moraes *et al.*, 2011), *P. adenoplylla* Wall. (Nguyen *et al.*, 2023a). This detail is also considered important in the classification of this genus. Particularly, the morphology of the lower epidermal cells of the leaves in this species differs significantly from those studied previously, its distinguished by having papillae cells and thin-walled. In addition, the paracytic stomata, which are common in the Rubiaceae family (Metcalf and Chalk, 1950), only find in the lower epidermic of this species. Moreover, the trichomes are present in the abaxial surface of leaves, which are present in *P. hoffmannseggiana*, *P. carthagenensis*, *P. deflexa* and *P. vellosiana*.

The anatomical characteristics of the stems of *Psychotria condorensis*, first documented here, exhibit similarities with studies of other *Psychotria* species, including the shape of the main vein bundle, the presence of crystalloid and starch bundles within the stem, as well as the type of wood pith within the main vein bundle of the stem. Additionally, the primary structure of the stem, a previously undescribed feature in the original literature, is described for the first time, revealing numerous single or multicellular hairs formed by elongated epidermal cells, although outward appearance makes them difficult to discern. The characteristics of the secondary stem indicate a wood vascular structure consistent with the research of Koek-Noorman (1977), with wood classified as type II, and analogous to the anatomical features of *P. deflexa*, *P. vellosiana*, *P. leiocarpa* (Marques *et al.*, 2015).

The presence of crystals in higher plants is common and linked to physical defense, oxalate removal from metabolism, calcium storage, and light regulation during photosynthesis in shade-grown plants (Franceschi and Nakata, 2005). The presence of crystals in the *Psychotria* species is important diagnostic characteristics for the genus (Moraes *et al.*, 2011). Raphide bundles are prevalent in the Rubioideae subfamily, including *Psychotria*, according to Lersten (1974), who also found druses to be common in South American *Psychotria* species. In this study, the raphide bundles was found not only in the midrib, leaf blade, petioles, but also in the young and mature stems.

This is the first time that the phytochemical of *Psychotria condorensis* has been reported in the study. The majority of constituents, including flavonoids, tanins, saponins, carbohydrates, coumarins, amino acids, were scientifically proven to display potent biological importance, and have beneficial impacts on treating diseases among others anti-cancer, antibacterial,

antioxidant, prevent diabetes, and additional biological functions (Banno *et al.*, 2004, Venugopala *et al.*, 2013; Ramamurthy and Sathiyadevi, 2017).

To formulate conservation strategies for the species, species identification, density assessment, and research on both macro and micro morphological characteristics are crucial. Apart from providing information for classification and phylogeny construction, plant morphology and anatomy elucidate the relationship between these structures and their physiological functions. The information provided by this study will contribute to establishing a foundation for conservation, propagation, and the potential development of medicinal resources for the species in the future.

ACKNOWLEDGMENTS

Nguyen Quoc Bao was funded by the Master, PhD Scholarship Program of Vingroup Innovation Foundation (VINIF), code [VINIF.2023.TS.010]. The authors would like to thank to Mr. Tran Song Hao, and all the staff of Con Dao National Park for their kindly supported during the field survey. We would like to thank you Dr. Pankaj Kumar for his kindly discussion. Sincere thanks also to the curators and staffs of the herbaria P, K, and VNM for their help in accessing specimens for this study.

LITERATURE CITED

- Andersson, L. 2002 Relationships and generic circumscriptions in the *Psychotria* complex (Rubiaceae, Psychotrieae). *Systematics and Geography of Plants* **72**: 167–202.
- Banno, N. B., Akihisa, T., Tokuda, H., Yasukawa, K., Higashihara, H., Ukiya, M. 2004 Triterpene acids from the leaves of *Perilla frutescens* and their anti-inflammatory and antitumor-promoting effects. *Biosci. Biotechnol. Biochem.* **68**(1): 85–90.
- Beentje, H. 2012 The Kew Plant Glossary, an illustrated dictionary of plant terms (revised edition). Kew: Royal Botanic Gardens, Kew Publishing. 160 pp.
- Benevides, P. J. C., Young, M. C. M., Bolzani, V. da S. 2005 Biological activities of constituents from *Psychotria spectabilis*. *Pharm. Biol.* **42**(8): 565–569.
- Bui, M.L., Tran, T.T.Q. 2013 Study of microscopic characteristics and chemical components of *Psychotria rubra* (Lour.) Poir. *Y Hoc TP. Ho Chi Minh* **18**(Supplement 1): 185–190.
- Chen, T., Taylor, C.M. 2011 *Psychotria*. In: Chen T., Taylor C.M., Swartz C. (eds.) *Flora of China*, vol 19. Science Press and Missouri Botanical Garden Press, Beijing and St. Louis, pp 294–301.
- Davis, A.P., Bridson, D., Jarvis C., Govaerts, R. 2001 The typification and characterization of the genus *Psychotria* L. (Rubiaceae). *Bot. J. Linn. Soc.* **135**(1): 35–42.
- de Oliveira, C.F., Oliveira V.B., Bobek, V.B., Rech, K. S., Betim F.C. M., Dias, J. de F.G., Zanin, S.M.W., Miguel, O.G., Miguel, M.D. 2020. Phytochemical and morpho-anatomical study of the vegetative organs of *Psychotria fractistipula* L.B.Sm., R.M. Klein & Delprete (Rubiaceae). *Braz. J. Pharm. Sci.* **56**: e18158



- Dwyer, J.D. 1980 Rubiaceae. In R.E. Woodson Jr., R.W. Schery & J. D. Dwyer (editors), Flora of Panama. Ann. Mo. Bot. Gard. **67**: 335–336.
- Franceschi, V.R., Nakata, P.A. 2005. Calcium oxalate in plants: Formation and function. Annu. Rev. Plant Biol. **56**(1): 41–71.
- Hamilton, C.W. 1989 A revision of Mesoamerican Psychotria subgenus Psychotria (Rubiaceae), part 1: introduction and species 1–16. Ann. Mo. Bot. Gard. **76**(1): 67–111.
- IUCN Standards and Petitions Committee 2019 Guidelines for Using the IUCN Red List Categories and Criteria. Version 14. Prepared by the Standards and Petitions Committee.
- Koek-Noorman, J. 1977 Systematische Holzanatomie einiger Rubiaceen. Ber. Deutsch. Bot. Ges. **90**(1): 183–190.
- Lersten, N.R. 1974 Morphology and distribution of colleters and crystals in relation to the taxonomy and bacterial leaf nodules in *Psychotria* Rubiaceae. Am. J. Bot. **61**(9): 973–981.
- Marques, J.B., Callado, C.H., Rabelo, G.R., Neto, S.J. da S., Cunha, M.da. 2015 Comparative wood anatomy of species of *Psychotria* L. (Rubiaceae) in Atlantic Rainforest remnants of Rio de Janeiro State, Brazil. Acta Bot. Bras. **29**(3): 433–444.
- Metcalf, C.R., Chalk, L. 1950. Anatomy of the dicotyledons, vol I. Oxford, Clarendon Press. 800 pp.
- Moraes, T.M. de S., Rabelo, G.R., Alexandrino, C.R., da Silva Neto, S.J., Cunha, M.D. 2011 Comparative leaf anatomy and micromorphology of *Psychotria* species (Rubiaceae) from the Atlantic Rainforest. Acta Bot. Bras. **25**(1): 178–190.
- Nepokroeff, M., Bremer, B., Sytsma, K.J. 1999 Reorganization of the Genus *Psychotria* and Tribe Psychotrieae (Rubiaceae) Inferred from ITS and *rcbL* Sequence Data. Syst. Bot. **24**(1): 5–27.
- Nguyen, K.P.P. 2007 Phương pháp cô lập hợp chất hữu cơ. Vietnam National University Ho Chi Minh Press, Ho Chi Minh city, 528 pp.
- Nguyen, Q.B., Pham, V.N., Quach, V.T.E., Truong, B.V., Pham, Q.T., Dang, V.S. 2023a Morphological, anatomical and distribution characteristics of *Psychotria adenophylla* Wall. In the southern islands of Vietnam. Thu Dau Mot University Journal of Science **6**(67): 24–33.
- Nguyen, Q.B., Quach, V.T.E., Huynh H.D., Pham, Q.T., Truong, B.V., Yahara, T., Tagane, S., Dang, V.S. 2023b A new species of *Psychotria* (Rubiaceae) from Bidoup – Nui Ba National Park, Vietnam. Phytotaxa **618**(2): 188–194.
- Nguyen, T.K.M. 2020 Kiểm nghiệm dược liệu. Vietnam National University Ho Chi Minh Press, Ho Chi Minh city, 212 pp.
- Pham, H.H. 2000 Cây cỏ Việt Nam - An Illustrated Flora of Vietnam, vol. 3. Youth Publishing House, Ho Chi Minh City. 999 pp.
- Pimenta, A.T.A., Uchôa, D.E.A., Braz-Filho, R., Silveira, E.R., Lima, M.A.S. 2011. Alkaloid and other chemical constituents from *Psychotria stachyoides* Benth. J. Braz. Chem. Soc. **22**(11): 2216–2219.
- Pitard, J. 1924 Rubiacées. In: H. Lecomte (ed.), Flore générale de l'Indo-Chine 3. Masson et Cie, Paris. pp. 20–442.
- Ramamurthy, V., Sathiyadevi, M. 2017 Preliminary phytochemical screening of methanol extract of *Indigofera trita* Linn. J. Plant Biochem. Physiol. **5**(2): 1000184.
- Taylor, C.M., Domínguez-Lincona, Ochoterena, H. 2007 A new species of *Psychotria* (Rubiaceae, Psychotrieae) from west-central Mexico. Novon **17**: 105–109.
- Tran, C.K. 1981 Microscopic techniques used in plant and medicinal research. Medical Publishing House, Hanoi, 150 pp.
- Tran, H.D.T., Nguyen, H.P.N., Nguyen, T.A., Mai, T.C., Duong, T.H. 2024 Isolation of some compounds from *Psychotria adenophylla* Wall. Ho Chi Minh city University of Education Journal of Science **21**(3): 376–380.
- Tran, N.N. 2005 Rubiaceae. In: Nguyen T.B. (eds.), Checklist of plants species of Vietnam. Agriculture Publishing House, Hanoi, 532 pp.
- Tran, P.H. 2018 Nguyên cứu về thực vật, thành phần hóa học và một số tác dụng sinh học của cây Hề Mọ (*Psychotria prainii* H. Lév.). PhD Thesis in Pharmacy. National Institute of Medicinal Materials, 248 pp.
- Turland, N.J., J.H. Wiersema, F.R. Barrie, W. Greuter, D.L. Hawksworth, P.S. Herendeen, S. Knapp, W.-H. Kusber, D.-Z. Li, K. Marhold, T.W. May, J. McNeill, A.M. Monro, J. Prado, M.J. Price and G.F. Smith (eds.) 2018 International Code of Nomenclature for algae, fungi, and plants (Shenzhen Code) adopted by the Nineteenth International Botanical Congress Shenzhen, China, July 2017. Regnum Vegetabile 159. Glashütten: Koeltz Botanical Books.
- Venugopala, K.N., Rashmi, V., Odhav, B. 2013 Review on Natural Coumarin Lead Compounds for Their Pharmacological Activity. Biomed Res. Int. **2013**: 963248.
- Vo, V.C. 2012 Từ điển cây thuốc Việt Nam, vol 1. Medical Publishing House, pp 90–100.

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